THE EFFECT OF LASER AND LASER PHOTOSENSITIZER COMBINATION ON LEISHMANIA TROPICA PROMASTIGOTES IN VITRO

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ABSTRACT

Cutaneous leishmaniasis is a common disease in Iraq, especially in the central part of the country. Several treatments have been suggested for this disease but none is completely effective and without side effects. Several research studies are focused on the development of alternative treatments.

The study was carried out in College of Medicine, Tikrit University, to show the effect of laser and laser photosensitiser (ultrameladinine) combination on Leishmania tropica promastigotes in vitro, after exposure for 3, 6 and 9 minutes. It was found that exposure to light from 5 mw lasers in the presence of ultrameladinine lead to destruction of L. tropica promastigotes in vitro. The effect of laser and laser photosensitiser was greater than laser light alone. Their effect on the parasite after exposure for 9 minutes was significantly greater than exposure for 3 and 6 minutes.

تأثير أشعة الليزر و ليزر التراميلادينين على اللشمانيا الجلدية البروماستيكوت في الزجاج (promastigotes)

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الخلاصة

اللشمانيا الجلدية هي من الأمراض الشائعة في العراق وخاصة في المنطقة الوسطى من البلاد.

وبدون أعراض جانبية المرض ولكن ليس فيهم علاج فعال وبصورة كاملة وبدون أعراض جانبية المديرة الدراسات البحثية تركزت لإيجاد العلاجات البديلة

أجريت الدراسة في كلية الطب- جامعة تكريت، لبيان تأثير أشعة الليزر وأشعة الليزر ما الترامية التراميلادينين على الطور البروماستيكوت (promastigots) للشمانيا الجلدية في الزجاج.

في الزجآج. أثبت أن التعرض لأشعة الليزر بوجود محلول التراميلادينين قد أعطى أفضل النتائج في قتل طفيلي اللشمانيا من أشعة الليزر وحده. وجدت أن تأثير هما عند التعرض لمدة (9) دقائق كانت أكثر من التعرض لمدة 3 و6 دقائق. وجدت أن التعرض من (5mw Laser) ليزر بوجود التراميلادينين يؤدي الى قتل الطفيلي في الزجاج.

INTRODUCTION

Laser is an acronym of light amplification by the stimulated emission of radiation and is advice that convert electrical energy in to light energy (1).

Laser beam have three special characteristics that differentiate them from ordinary light. Monochmaic (all of the same color or wavelength), collimated (emitted in a narrow parallel beam) and coherent (all the light waves are in synchrony). As a result of collimation there is a very little divergence over long distances (2).

When laser beam encounters matter, photons are either reflected, transmitted, scattered or absorbed. A portion of the beam might be reflected by being back scattered, transmitted without effect on the tissue or absorbed. Absorption of laser lights depends on many parameters like wavelength and the types of tissue. There are three basic types of effect on living tissue such as photothermat (3), photochemical (2) and photo-caustic (2). In several branches of medicine, laser is used as therapeutic agents such as in ophthalmology, dermatology, gynecology and surgery (4).

There have been several conducted reports on the ability of laser to kill the microorganism in vitro. Mathews and Sistrom (5) demonstrated that a toulidine blue-sensitized colorless mutant of Sarcina lutea could be killed by polychromatic light from tungsten and fluorescent lamps. Wilson (6, 7 & 8) reported that various toludine blue O (TBO) sensitized oral bacteria like Porphyromonas gingivalis, Fusobacterium mucleatum and Actinobacillus actinomycetemomitans could be killed by low doses of helium-neon laser light. Macmillan et al., (9) found that gram positive Sarcina lutea and gram negative Escherichia coli and Pseudomonas aeruginasa could be killed by helium-neon light in the presence of the dye. Al-Obaidi (10) found that providine-iodine sensitized Staphylococcus aureus could be killed by helium-neon laser.

Leishmania parasites are thermosensitive. In in vitro study, L. tropica multiplied best at 35 C and was completely eliminated at 37 C. In view of this feature both heat and cold treatment have been tried. In Iraq, infrared heat was used to raise the temperature of CL lesions to 55 C for 5 minutes, and all lesions healed in 5 to 6 weeks (11). Mutinga and Mingola successfully treated three cases of acute CL by combined ultraviolet light and infrared therapy (12).

Although laser has been used for treatment of several skin diseases since 1970, but there are controversies about the effect of laser therapy on the processes of wound healing, connective tissue repair and collagen biosynthesis. Rackcheev, et al., (13), found complete clinical cure in patients suffering from skin leishmaniasis with an argon laser. It has been shown that Helium-Neon low energy laser at wave length 630 nm is effective in wound healing (14). Babejev, et al., (15), used carbon dioxide laser to vaporize the local lesions caused by cutaneous leishmaniasis. They found that the treatment reduces the management of patient at 1.5 times and is followed by satisfactory aesthetic outcomes. Eissa, et al., (16) investigated the mechanism of action and efficacy of CO2 laser rays and hypertonic sodium chloride (NaCl) with different concentration in treatment of cutaneous leishmaniasis. They concluded that CO2 laser then 7% NaCl are good modalities for cutaneous leishmaniasis treatment. Asilian et al., (17) found that the CO(2) laser was more effective in treating cutaneous leishmaniasis than glucantime (1.12 times), had fewer side effects and resulted in a shorter healing time. In Isfahan, Iran, Asilian, et al., (18) showed that carbon dioxide laser radiation is highly effective to vaporize local cutaneous leishmaniasis in 24 patients with lupoid cutaneous leishmaniasis.

The study was conducted to show the effect of laser and laserphotosensitiser combination on Leishmania tropica in vitro.

MATERIALS AND METHODS

Laser: The laser was diode gas laser with a measured output of 5 mw (Laser Becaon, INC., Michigan, USA). This emits light with a wavelength of 630 nm in a collimated beam with a diameter 1.3 mm.

Photosensitiser: Ultrameladinine (Memphis Co. for Pharm. And Chemical Ind. A.R.E.) was used as a photosensitiser.

Parasite: Leishmania tropica promastigotes was obtained from the Leishmania unit at the Research Center/ Nahrin College of Medicine.

Experimental design: Attempt was made for the killing of L. tropica promastigotes by using laser light alone and in combination with ultrameladinine as photosensitiser.

Promastigotes was grown in liquid media at 26 ^oC for six days. After 6 days of incubation heavy growth was obtained. On the day of experiment, promastigotes were harvested and diluted to 1×10^6 promastigotes /ml, then the aliquots (100 ml) of the suspension was transferred to a sterile test tube and an equal volume of ultrameladinine solution was added to each tube to give a final concentration equal to 0.002 mg/ml. The test tube was placed on a magnetic stirrer and exposed to laser light for 3, 6 and 9 minutes. Control tubes containing the promastigotes suspension and saline solution instead of ultrameladinine were treated as before, to determine the effect of laser radiation alone on promastigotes viability. A further four tubes as those described above, were prepared, were not exposed to laser light. Each run was done in duplicate. They were incubated at 26 C for 5 days (7 &10).

On the next five days the cultures were counted a 1:10 dilution in saline together with the appropriate dye was prepared. The dye for promastigotes was 0.4 % trypan blue. The promastigotes permeable to the blue dye are dead while viable ones exclude the dye. The chamber of Neubaurs slide is used for counting the promastigotes (19).

The total number per ml= No. counted x 10 (number in mm) x 1000 (number in ml) x 10 9 dilution factor.

Growth index (GI) % = No. of treated promastigotes divided by number of untreated promastigotes multiply by 100.

Statistical analysis: Analysis of variance test was performed to show significant difference between groups.

RESULTS

The number of live L. tropica promastigotes after 3, 6 and 9 minutes exposure to laser and laser-photosensitiser is shown in table (1). It is illustrated that the number of promastigotes on day 5 after exposure were 22.75, 16.88 and 24.9×10^6 in laser, laser-photosensitiser combination and control groups respectively.

It was found that on 6 minutes of exposure, the number of promastigotes on day 5 were 22.65, 5.85 and 24.9⁶ in laser, laser-photosensitiser combination and control, respectively. On 9 minutes exposure, it was obvious that laser-photosensitiser had more significant effect than laser light alone and the number

of promastigotes on day 5 after exposure were 15.43, 0.85 and 24.9×10^6 in laser, laser-photosensitiser combination and control groups, respectively.

The growth index of L. tropica in vitro, illustrated in table (2). It was found that laser-photosensitiser exposure lead to decrease in growth index from 100% at day 0 to 3% at day five after exposure, while growth index after exposure to laser light alone was 61%. Statistically there was significant difference between them (P<0.01).

	Joinastigotes invitro.					
	Light	Day 1	Day 2	Day 3	Day 4	Day 5
Incubation						
Time of						
exposure						
(Minutes)						
3	L	2.7	12.6	24.0	29.6	23.0
	L-U	2.4	8.3	18.2	21.6	17.1
6	L	2.5	12.5	23.6	20.5	22.0
	L-U	2.0	5.2	7.0	8.8	7.0
9	L	2.2	11.6	19.7	19.6	22.2
	L-U	2.0	4.2	3.6	3.0	1.8
Control		2.6	6.44	21.35	32.12	30.65

Table 1. Effect of laser (L) and laser-ultrameladinin (L-U) on L. tropica promastigotes invitro.

Table 2. The effect of laser (L) and laser-ultrameladinin (L-U) combination on
growth of L. tropica promastigotes in vitro on day 5 of exposure.

Time of exposure	Laser	Laser-Ultrameladinin		
(minutes)				
3	91.36	67.79		
6	62.57	14.7		
9	61.08	3.41		

DISCUSSION

Although pentavalent antimony is the treatment of choice for healing the lesions of cutaneous leishmaniasis, but it has a high incidence of side effects. In general, adverse reactions are expected in patients with liver and renal impairment, cardiac arrythmias and prolonged QT intervals, in small children, in breast feeding women and in pregnant and obese elderly and immunocompromoised patients (20). In addition to that lesion of cutaneous leishmaniasis is usually resistant to the conventional therapies for leishmaniasis and it may persist and spread slowly for many years (21). Therefore it is worthwhile to find alternative treatment for destroying the parasite and healing of lesion of cutaneous leishmaniasis.

In literature review, we did not come across the application of laser and photosensitiser combination for treatment of lesion of cutaneous leishmaniasis. In the present preliminary study the effect of laser and laser photosensitiser combination was applied against promastigote stage of the parasite in vitro. Iraqi Journal of Veterinary Sciences, Vol. 20, No.2, 2006 (117-122)

The results of this study have demonstrated that, exposure to light from 5mw laser with ultrameladinine (methoxsalen) resulted in significant decline in promastigotes number, and at 9 minutes exposure of their combination lead to complete destruction of promastigotes number. This approach of killing promastigotes is photochemical method (3), which was chosen on the basis of literature report and criteria which used for killing of microorganisms (9 &10).

It is concluded that laser photosensitiser combination had greater efficacy for killing of promastigote stage of parasite than laser light alone. As the results of in vitro study is not applicable for treatment of cutaneous leishmaniasis in vivo. It is recommended to carry on further studies to use laser-photosensitiser combination for treatment of skin lesion of cutaneous leishmaniasis in vivo and a trial of its use for vaccine production against this disease.

REFERENCES

- 1. Genack AZ, Drake, JM. Scattering for super-radiation. Nature 1994; 400-401.
- 2. Nayantara B, Keith MB, Anderson RR. Comparative study of different laser systems. Fer. Steri 1994; 61: 581-591.
- 3. Hits CB. Understanding laser technology. Pann. Well, USA; 1984.
- 4. Goldman MP, Fitzpatrick RE. Laser treatment of scars. Dermatol Surg 1995; 21: 685-687.
- 5. Mathews MM, Sistrom WR. The function of carotenoid pigments of Sarcina lutea. Arch. Microbiol 1960; 35: 139-146.
- 6. Wilson M, Dobsan, J, Sarkar S. Sensitization of reriodentopathogenic bacteria to killing by light from a low power laser. Oral Microbiol Immunol 1993; 8: 182-187.
- 7. Wilson M. Photolysis of oral bacteria and its potential use in treatment of caries and periodontal disease. J Appl Bacteriol 1993; 75: 299-306.
- 8. Spikes JD, Jori G. Photodynamic therapy of tumors and other diseases using porphyrins. Laser Med Sci 1987; 2: 3-15.
- 9. Macmillan JD, Maxwell WA, Chichester Co. Lethal photosensitization and micro-organisms with light from a continous wave gas laser. Photochem Photobiol 1966; 5: 555-565.
- 10. Al-Obaidi HS. A study on Staphylococcus aureus strains isolated from wounds and their response to various antimicrobial agents and He-Ne laser radiation. M Sc thesis, College of Medicine, Tikrit University 1996.
- 11. Junaid AJN. Treatment of cutaneous leishmaniasis with infra-red heat. Int J Dermatol 1986; 25: 470-472.
- 12. Mutinga MJ, Mingola EN. Alternate treatment of cutaneous leishmaniasis. East Afr Med J 1974; 51: 68.
- 13. Rakcheev AP, Chistiakova IA, Kamennykh PV. The successful treatment of cutaneous leishmaniasis with argon laser. Vestn Dermatol Venerol 1989; 12: 53-55.
- 14. Tak-Zare N, Sarkandy M, Foroutan MR, Yarmohammadi K, Tak-Zare S, Tak-Zare AR. The effect of Helium-Neon low energy laser Beam on the healing of injured tissues. Irn J Plastic Reconstructive Surg 1999; 2: 45-49.
- 15. Babajev KB, Babajev OG, Korepanov VI. Treatment of cutaneous leishmaniasis using a carbon dioxide laser. Bull World Health Organ 1991; 69: 103-106.
- 16. Eissa MM, Soliman AS, Nassar SO. Ultrastructural and immunological features of experimental cutaneous leishmaniasis after treatment with

intralesional hypertonic sodium chloride and CO2 laser rays. J Egypt Soc Parasitol 2003; 33; 329-352.

- 17. Asilian A, Sharif, A, Faghihi G. Enshaeich Sh, Shariati F, Siadat AH. Evaluation of Co laser efficacy in the treatment of cutaneous leishmaniasis. Int J Dermatol 2004; 43: 736-738.
- Asilian A, Iraji F, Hedaiti HR, Sidat, AH, Enshaieh S. Carbon dioxide laser for the treatment of lupoid cutaneous leishmaniasis (LCL): A case series of 24 patients. Dermatology Online J 2006;12: 3-9.
- Najim RA, Sharquie KE, Farjou IB. Zinc sulfate in the treatment of cutaneous leishmaniasis an in vitro and animal study. Men Inst Oswaldo Ciuz 1998; 93: 831-837.
- 20. Markle WH, Makhoul K. Cutaneous leishmaniasis: Recognition and Treatment. Amer Family Physician 2004; 69: 1339-1398 (from internet).
- Vega Lopez F, Hay RJ. Parasitic worms and protozoa, In: Breathnach, S, Cox N. et al. Rook textbook of dermatology. 7 th. Ed London, Blackwell Science 2004; Vol. 2: pp: 32, 35, & 42.