Study of the Biological Activity of Aqueous Extract of *Cuminum cyminum* L. and *Hibiscus sabdariffa* L. and Detection of Some Active Groups in them

دراسة الفعالية الحيوية للمستخلصات المائية لنباتي الكمون والكجرات والكشف عن بعض المجاميع الفعالة

فيها

Hussein Kadhem Abdul Hussein (PhD)*
Najeh Hashem Kadhem (MSc)**
Zuhair Hamid Abbod (MSc)**
(*): Department of Chemistry, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Biology, College of Science, University of Karbala, Iraq.
(*):Department of Chemistry, College of Science, E-mail: headm2000@yahoo.com.

Abstract:

The studies about the effect of plant extract against different types of bacteria are still one of the most important fields of researches because they are available, cheep, safe, and easy to use by not professional populations. In this work, two plant extracts (Hibiscus sabdariffa) and Cuminum cyminum) were tested for there possible biological activity against bacteria. Boiled aqueous extract of both plants at the following concentrations (1, 5, 10, and 20%) were used after cooling. Agar well diffusion was used to examine the biological activity of each extract and the results expressed as zone of inhibition in (mm). The bacteria used in this study are (Gram positive: Streptococcus faecalis and Staphylococcus aureus) and (Gram negative: Pseudomonas pyogens, Escherichia coli, and Klebsiella pneumoniae). The tests for functional groups that can be extracted by water were carried out. Hence Alkaloids, Saponines, Tannins, Glycosides were screened using a suitable method while oils noticed as upper layer (if present). At 1% concentration, there are no detectable inhibition zones of both aqueous extracts. The inhibition zones appeared at 5% and further inhibition showed as concentration increased. In general, the aqueous extract of Hibiscus sabdariffa has shown more biological activity than the corresponding concentration of *Cuminum cyminum*. The lowest activity was shown against Escherichia coli bacteria while the other bacteria showed different zone diameters as a response for the same concentration. In Cuminum cyminum there are detectable amounts of essential oils, glycosides and high amounts of tannins while alkaloids and saponines are not detectable using the described methods. In Hibiscus sabdariffa, there is a detectable amount of saponines, glycosides, and high amounts of tannins while there are no positive results for essential oils and alkaloids. In conclusion, the aqueous extracts of H. sabdariffa and Cuminum cyminum have antibacterial activity at concentrations $\geq 5\%$. Hibiscus sabdariffa has shown more biological activity than the corresponding concentration of Cuminum cyminum. This fact may be due, in part, to the presence of saponines in Hibiscus sabdariffa. The results can be explained through the presence of different active substances in the aqueous extract of both plants.

<u>Keywords:</u> Biological activity, Aqueous extracts, *Cuminum cyminum L., Hibiscus sabdariffa L.*, Alkaloids, Tannins, Saponines, Glycosides, oils.

الخلاصة

لأزالت در اسة الفعالية البايولوجية للمستخلصات النباتية واحدة من الحقول البحثية المهمة كون هذه المستخلصات النباتية لنباتي الكجرات (hibiscus الاستعمال من قبل الناس غير المتخصصين. في هذا البحث تمت در اسة الفعالية البايولوجية للمستخلصات المائية لنباتي الكجرات (hibiscus التعمل من قبل الناس غير المتخصصين. في هذا البحث تمت در اسة الفعالية البايولوجية للمستخلصات المائية لنباتي الكجرات (*Cuminum cyminum*) والبحث عن بعض المواد الفعالة فيها. استعملت المستخلصات المائية المغلية لكل مستخلص وحسبت (*sabdariffa التراكيز الآتية (١%، ٥%، ١ ١%، و ٢٠%) بعد تبري*دها. واستعملت طريقة الحفر بالاكار لاختبار الفعالية البايولوجية لكل مستخلص وحسبت *Streptococcus من*فقة التثبيط بوحدة المليمتر. اختبرت فعالية المستخلصات ضد البكتيريا الآتية الموجبة لصبغة گرام (*Staphylococcus geee الإكار لاختب*ار الفعالية المستخلص وحسبت *و العاوية بي قل من منطقة التثبيط بوحدة المليمتر. اختبرت فعالي المستخلصات ضد البكتيريا الآتية الموجبة لصبغة گرام (Staphylococcus geee العود العالية وحسبت <i>و العاية بي و وحدة و staphylococcus و وحدة من و مع من و حود المجاميع الفيا لا ي و وحدة من عدم على معاح مي الغيا في علم الاختبار الكرفي و تلاين المودين و مع أمينية و سبعة و والصابونينات و التانينات <i>و التاين في و معامة و و الحرينيا و الفود و البي و وجدة من و و و و و ماي ما لاختبار الكشف عن و و و المود و و و و و ماي ما معد ما موجب الموجبة لمود من العورينات و التانينات <i>و التانيا و و و الحري و ما و الخوي و و و المود و و الكيو و و و و و و المود و و العابي و و و و و المود و ما و مي ما و التانيات <i>و التانيا تر و العابي و الكليوسيدا و التانيا و اللاي و مستخلص و الليو و و و الكيو و و الكليو و و الكليو و و و العربي و الخري و ما معن الغراب المود و ما معن و ما معن مود و المود مع ما مود المود مع ما مود و مع ما مود و ما ما ما معن عدم ما مود و ما ما و العابو و و الكليو و و م المود و ما ما و ما ما و و و ما ما و و ما ما ما و ما معرم ما ما و المور و ما م و الصابونينات و الكلايكوسيدات الما معرف المود مع المود مع ما عمون الكمون الكليون و التانيو و مع المود و و ما ما و الفود و ما ما و و ما ما و لاهور و ما ما و ما ما و و ما ما و ما مود ما ما ما ما و ما ما و ما مون و مما ما الكمون و ما ما*

مفاتيح الكلمات: الفعالية البايولوجية، المستخلصات النباتية، الكجرات (Hibiscus sabdariffa)، الكمون (Cuminum cyminum) القلويدات، التانينات، الصابونينات، الكلايكوسيدات، الزيوت

Introduction:

A lot of antimicrobial screening evaluations have been published based on the traditional use of Chinese, African, and Asian plant drugs during this period. The studies about the effect of plant extract against different types of bacteria are still one of the most important fields of researches $^{(1, 2, 3, 4)}$.

Many new antibacterial agents submitted to analysis by the American food and drug agency (FDA) and some of them were natural products while others were semisynthetic products modeled on a natural product lead ⁽⁵⁾.

Hibiscus sabdariffa L. (Rosella) water extract is a local soft drink material and medicinal herb. The *Hibiscus sabdariffa* extract consider to be save knowing that LD(50) of roselle calyx extract. The LD(50) was found to be above 5000 mg kg^{-1 (6)}. Though the average consumption of 150-180 mg/kg per day appears safe, the extract should be taken with caution bearing in mind that higher doses could affect the liver ⁽⁷⁾. *Hibiscus sabdariffa* exhibits antihypertensive and cardioprotective effects in vivo and supports the public belief that *Hibiscus sabdariffa* may be a useful antihypertensive agent ^(6, 8, 9, 10). In one study ⁽¹¹⁾, the results suggested that *Hibiscus* extract blocks adipogenesis, in part, by its suppression on the expression of adipogenic transcription factors ⁽¹¹⁾. These results suggest that *Hibiscus sabdariffa* L. inhibits serum lipids and shows an antiatherosclerotic activity ⁽¹²⁾.

Administration of a crude extract of *Hibiscus sabdariffa* have protected erythrocytes against lipid peroxidation ⁽¹³⁾. Water extract of the dried flowers of *Hibiscus sabdariffa* L. reduces the hepatotoxicity caused by different agents ^(14, 15, 16). The results indicate that rosella has antimutagenic activity ⁽¹⁷⁾. The data suggested that *Hibiscus* protocatechuic acid is an apoptosis inducer in human leukemia cells ⁽¹⁸⁾. These results indicate that *Hibiscus* protocatechuic acid (PCA), a phenolic acid isolated from *Hibiscus sabdariffa* L., possesses potential as a cancer chemopreventive agent against tumor promotion ⁽¹⁹⁾.

Cuminum cyminum L. (Cumin) is a small annual herb native to the Mediterranean region. India and Iran are the largest cumin exporters. The valued portion of the plant is the dried fruit called cumin seed, which is esteemed as a condiment. The odor and flavor of cumin is derived largely from the essential oil, which contains cuminaldehyde as the main constituent. Other ingredients of the oil are dihydrocuminaldehyde, d, 1-pinene, d--pinene, para-cymene, -pinene, dipentene, and cuminyl alcohol. The dried seeds of cumin have 2.5 to 5 percent essential oil on a dry weight basis as obtained by steam distillation. As a medicinal plant, cumin has been utilized as an antispasmodic, carminative, sedative, and stimulant. Cumin oil has been reported to has antibacterial activity (1.8-130). Distinct phototoxic effects have been reported from undiluted cumin oil (8.2-79) Cumin is generally recognized as safe for human consumption as a spice/flavoring and plant extract ⁽²⁰⁾. The main essential oils of *Cuminum* L. oil were p-mentha-1,4-dien-7-al, cumin aldehyde, gamma-terpinene, and beta-pinene⁽²¹⁾.

The essential oils extracted from the seeds of *Cuminum cyminum*, have been studied for antibacterial activity against different eight pathogenic bacteria, causing infections in the human body. It has been found that the oil of *Carum capticum* is very effective against all tested bacteria. The oil of *Carum cyminum* and *Anethum graveolens* also gave effective against all tested bacteria⁽²²⁾.

The present study has been designed to identify antibacterial activity in extracts obtained from against type cultures and organisms that were isolated form various infections. Some tests for active groups in these aqueous plant extract also studied.

Materials and Methods

1. Collection and Extraction of Plant Material:

Whole dried calyces of the flower of *Hibiscus sabdariffa* and dried seeds of *Cuminum cyminum L*. were collected from the market and classified by expert taxonomist. The plant materials were carefully sorted out to remove any debris or possible contaminating substances. Then they were pulverized in a mill. The following quantities of the dried plant parts (1, 5, 10, and 20 grams) were then covered with 100 milliliters of distilled water and thoroughly extracted by boiling for 15 minutes. The evaporated water was substituted by adding an extra volume of distilled water to obtain a final volume of 100 milliliters of filtered extract.

2. Identification of Active groups in aqueous plant extract:

The tests that can be carried out to screen functional groups that can be extracted by water were carried out. While those tests that required alcoholic extract were not performed because this work deals with water soluble compounds in both plants. Hence tests about flavones, coumarins, and terpenes were not done while oils noticed as upper layer (if present). Alkaloids measured using Dragendroff's reagent according to Harborn (1973) method ⁽²³⁾. Saponines screened by two methods. The first method was carried out by mercuric chloride method. The second method by vigorous mixing of a tube containing the extract; if suds formed and keep on for 5-10 minutes this indicates the presence of saponines ⁽²⁴⁾. Tannins screened by ferric chloride method as described by Harborn (1984) ⁽²⁵⁾. Glycosides screened by Fehling's reagent as described by Adeday *et al* (2001) ⁽²⁶⁾.

3. Antibacterial Activity:

The agar-well diffusion method (Perez *et al* 1990) ⁽²⁷⁾ was used to screen the antibacterial activity of each extract. Aliquots of 20mL of Müeller Hinton agar (Oxoid[®], Basingstoke, Hampshire, England) were poured into 9 cm Petri dishes in order to evaluate bacterial growth. The agar inoculated with 100 μ L of 18-24 hour nutrient broth cultures of the following bacteria (Gram positive: *Streptococcus faecalis* and *Staphylococcus aureus*) and (Gram negative: *Pseudomonas pyogens, Escherichia coli, and Klebsiella pneumoniae*). Four holes, equidistant from each other and from the edge of the plate, were bored into the solidified seeded agar using sterile glass borers (8 mm in diameter). The plates were incubated at 37°C for 30 minutes before introducing 50 μ L of each extract (at 1%, 5%, 10%, and 20%) into each of two holes using micropipette. Holes containing distilled water alone were included on each plate as control. The plates were refrigerated at 4°C for 30 minutes to allow for diffusion before incubating at 37 °C for 24 hours. The produced zones of inhibition were measured for each organisms screened.

Results:

Antibacterial activity of *H.sabdariffa* and *Cuminum cyminum* was measured as diameter of the inhibition zone (mm) as shown in Tables (1 and 2). The results of both plants showed that the inhibitions of bacterial growth are concentration dependent. Table (1): Biological activity of the aqueous extract of *Hibiscus sabdariffa* L.

Bacteria type	Zone of Inhibition (mm)			
	1%	5%	10%	20%
Streptococcus faecalis	0	16	18	22
Staphylococcus aureus	0	24	24	24
Pseudomonas pyogens	0	14	18	26
Escherichia coli	0	14	18	18
Klebsiella pneumonia	0	3	18	20

expressed as zone of inhibition (mm) using agar well diffusion method.

Zone of minorition (min) using agar wen unrusion method.							
Bacteria type	Zone of Inhibition (mm)						
	1%	5%	10%	20%			
Streptococcus faecalis	0	14	18	18			
Staphylococcus aureus	0	12	12	18			
Pseudomonas pyogens	0	12	12	18			
Escherichia coli	0	3	3	14			
Klebsiella pneumonia	0	3	16	20			

 Table (2): Biological activity of the aqueous extract of *Cuminum cyminum* L. expressed as zone of inhibition (mm) using agar well diffusion method.

At 1% concentration, there are no detectable inhibition zones of both aqueous extracts. The inhibition zones appeared at 5% and further inhibition showed as concentration increased. In general, the aqueous extract of *H.sabdariffa* has shown more biological activity than the corresponding concentration of *Cuminum cyminum*. The lowest activity was shown against *E.coli* bacteria while the other bacteria showed different zone diameters as a response for the same concentration.

The active components were studied in 10% of aqueous plant extract to detect the active components in both plants (Tables (3)).

Table (3): Biological activity of the aqueous extract of *Cuminum cyminum* L. expressed aszone of inhibition (mm) using agar well diffusion method.

	Presence of detectable active components in 10% of Plant Extract						
Plant Extract	resence of detectable active components in 10% of Flant Extract						
	Essential Oils	Alkaloids	Tannins	Glycosides	Saponines		
Cumin	+	-	++	+	-		
Sabdhariffa	-	-	++	+	+		

In *Cuminum cyminum* there are detectable amounts of essential oils, glycosides and high amounts of tannins While alkaloids and saponines are not detectable using the described methods. In *H.sabdariffa*, there is a detectable amount of saponines, glycosides, and high amounts of tannins while there are no positive results for essential oils and alkaloids.

Discussion:

The search for antibiotics in plant extract still an attractive field of study because they are available, cheep, safe, and easy to use by unspecialized populations ^(28, 29, 30).

Gram-negative organisms tend to have a higher intrinsic resistance to most antimicrobial agents. In the study by Ndukwe *et al* (2005) ⁽¹⁾ the Gram-negative organisms were less susceptible than Gram-positive organisms to the activity of the different plant extracts. Antibacterial activity, determined with the agar diffusion method, was observed against Gram-positive and Gram-negative bacterial species in this study. The lower activity was observed against bacteria belonging to the genus *Pseudomonas*. These results suggested the potential use of the above essential oils for the control of bacterial diseases ⁽²¹⁾. The essential oil of different plant sources showed different antibacterial activity ^(31, 32), and many types of bacteria did not affected by essential oils of some plants ⁽³¹⁾.

Hibiscus protocatechuic acid (PCA), a phenolic compound found in the dried flowers of *Hibiscus sabdariffa L*. (Malvaceae), was demonstrated to have an antioxidant effect in vitro and *in vivo*, and an antitumor property in previous study ⁽¹⁵⁾. These results indicate that protocatechuic acid, a polyphenolic compound from *Hibiscus sabdariffa* L. protects against tert-butylhydroperoxide-induced hepatotoxicity by its antioxidant ^(33, 34) and anti-inflammatory characteristics accompanied by blocking of stress signal transduction ⁽³⁴⁾.

A roselle (Hibiscus sabdariffa Linn.) tea extract was found to have high inhibitory activity against porcine pancreatic α -amylase ⁽³⁵⁾ and the crude hydroalcoholic extract showed in vitro an appreciable enzyme-inhibiting activity towards the Angiotensin I Converting Enzyme (ACE), attributable to flavones, but weak inhibiting activities towards elastase, trypsin and alpha-chymotrypsin. The angioprotective activity in vivo, also important, was due to flavones and anthocyanins ⁽³⁶⁾. This ability to inhibit enzyme activity may be the reason about the antibacterial activity (i.e. may inhibit some enzymes necessary for bacterial biological activity. Anthocyanins are natural plant dyes present as glycosides and water soluble. The concentration of anthocyanins in the cold extract of *H.sabdariffa* found to be 5% while the hot extract of H. sabdariffa contained the second highest concentration of anthocyanins (31%). The heat aided the extraction of the anthocyanins in H. sabdariffa, despite their susceptibility to rapid loss of color because of the acidified methanol used for extraction. It is recommended that extracts of both H. sabdariffa great commercial value as natural colorants in Nigeria ⁽³⁷⁾. Aqueous extracts of the calyces are concentrated with anthocyanins ⁽³⁸⁾. *H. sabdariffa* contain different anthocyanins in addition to their colourful characteristics possess antioxidant properties ^(39, 40). *Hibiscus sabdariffa* also contain lignin (41)

Hibiscus sabdariffa extract had an inhibitory effect on yeast induced pyrexia. Among the phytoconstituents found in both plants, flavanoids, polysaccharides, and organic acids may be mainly responsible for their pharmacological activities ⁽⁴²⁾.

Raw polysaccharides were isolated from the flowers of *Hibiscus sabdariffa* L. and fractionated by ion exchange chromatography into one neutral and three acidic subfractions. Raw polysaccharides and all acidic subfractions caused a strong induction of proliferation of human keratinocytes of up to 40 %, while the neutral polymers were ineffective. While mitochondrial activity was not influenced, raw polysaccharides induced early differentiation of primary natural human keratinocytes ⁽⁴³⁾. The neutral polysaccharides (HIB 1 and 2) are composed of arabinans and arabinogalactans of low relative molecular mass ⁽⁴⁴⁾.

The results also indicated that hibiscus contained low levels of aluminum and greater amounts of iron and copper ⁽⁴⁵⁾. The possible effect of these ions in the extract on biological activity needs more investigation.

Among the 49 compounds of volatile oil was extracted from *Cuminum cyminum* L. identified, there were 16 hydrocarbons and 32 oxygenated compounds. The main components were cuminal and safranal (accounting for 32.26% and 24.46% respectively in the components identified). The other nine compounds with contents all over 1% were monterpenes, sesquiterpenes, aromatic aldehydes and aromatic oxides etc. The other components with relatively small amounts were chiefly terpenes, terpenols, terpenals, terpenones, terpene esters and aromatic compounds ⁽⁴⁶⁾. From the water-soluble portion of the methanol extract of cumin (fruit of *Cuminum cyminum* L.), which has been used as a spice and medicine since antiquity, sixteen monoterpenoid glucosides, including twelve new compounds, were isolated. Their structures were clarified by spectral investigation ⁽⁴⁷⁾. Many spices including cumin were found to contain the following elements: Mg, Al, Si, P, S, Cl, K, Ca, Ti, Mn, Fe, Cu, and Zn, with varying concentrations. Mutagenic studies using different *Salmonella typhimurium* strains showed a very weak oxidative mutagenic action has been revealed by cumin ⁽⁴⁸⁾. Ethanolic extracts of *Cuminum cyminum* L. expressed minimum

inhibitory concentration (MIC90) values of 0.075 mg/mL. The results showed a significant *in vitro* effect of plant extracts against *H. pylori* that could be considered a valuable support in the treatment of the infection and may contribute to the development of new and safe agents for inclusion in anti-*H. pylori* regimens ⁽⁴⁹⁾.

In conclusion, the noticed biological activity in the present work showed that the activity depends on the chemical constituents of each plant extract and the activity is dependent on the concentration of the water soluble chemicals from each plant.

References:

1. Ndukwe KC, Okeke IN, Lamikanra A, *et al* (2005): Antibacterial activity of aqueous extracts of selected chewing sticks. J Contemp Dent Pract. Aug 15;6(3):86-94.

- Sagdic O, Ozkan G, Ozcan M, Ozcelik S. (2005): A study on inhibitory effects of Sigla tree (Liquidambar orientalis Mill. var. orientalis) storax against several bacteria. <u>Phytother</u> <u>Res.</u> Jun;19(6):549-51.
- 3. McGaw LJ, Gehring R, Katsoulis L, Eloff JN.(2005): Is the use of Gunnera perpensa extracts in endometritis related to antibacterial activity? <u>Onderstepoort J Vet Res.</u> Jun;72(2):129-34.
- 4. Zhao CC, Shao JH, Li X, Xu J, Zhang P. (2005): Antimicrobial constituents from fruits of Ailanthus altissima SWINGLE. <u>Arch Pharm Res.</u> Oct;28(10):1147-51.
- 5. Cragg GM, Newman DJ & Snader KM (1997): Natural products in drug discovery and development. *Journal of Natural Products*, 60: 52-60.
- 6. Onyenekwe PC, Ajani EO, Ameh DA, Gamaniel KS.(1999): Antihypertensive effect of roselle (Hibiscus sabdariffa) calyx infusion in spontaneously hypertensive rats and a comparison of its toxicity with that in Wistar rats. <u>Cell Biochem Funct</u>. Sep;17(3):199-206.
- 7. Akindahunsi AA, Olaleye MT. (2003): Toxicological investigation of aqueous-methanolic extract of the calyces of Hibiscus sabdariffa L. J Ethnopharmacol. Nov;89(1):161-4.
- 8. Odigie IP, Ettarh RR, Adigun SA. (2003): Chronic administration of aqueous extract of Hibiscus sabdariffa attenuates hypertension and reverses cardiac hypertrophy in 2K-1C hypertensive rats. J Ethnopharmacol. Jun;86(2-3):181-5.
- Herrera-Arellano, S. Flores-Romero, M. A. Chavez-Soto (2004): Effectiveness and tolerability of a standardized extract from Hibiscus sabdariffa in patients with mild to moderate hypertension: a controlled and randomized clinical trial. <u>J. Tortoriello.</u> <u>Phytomed.</u> 11, 374 – 382.
- 10. Adegunloye B. J., Omoruyi J. O., Ajabonna O. P., *et al* (1996): Mechanisms of the blood pressure lowering effect of the calyx extract of Hibiscus sabdariffa in rats. <u>Afr. J. Med.</u> <u>Sci</u>. 25, 235 238.
- 11.Kim MS, Kim JK, Kim HJ, Moon SR, *et al* (2003): Hibiscus extract inhibits the lipid droplet accumulation and adipogenic transcription factors expression of 3T3-L1 preadipocytes. J Altern Complement Med. Aug;9(4):499-504.
- 12.Chen CC, Hsu JD, Wang SF, *et al* (2003): Hibiscus sabdariffa extract inhibits the development of atherosclerosis in cholesterol-fed rabbits. J Agric Food Chem. Aug 27;51(18):5472-7.
- 13.Suboh SM, Bilto YY, Aburjai TA. (2004): Protective effects of selected medicinal plants against protein degradation, lipid peroxidation and deformability loss of oxidatively stressed human erythrocytes. <u>Phytother Res</u>. Apr;18(4):280-4.

- 14.Ali BH, Mousa HM, El-Mougy S. (2003): The effect of a water extract and anthocyanins of hibiscus sabdariffa L on paracetamol-induced hepatoxicity in rats. <u>Phytother Res.</u> Jan;17(1):56-9.
- 15.Lin WL, Hsieh YJ, Chou FP, *et al* (2003): Hibiscus protocatechuic acid inhibits lipopolysaccharide-induced rat hepatic damage. Arch Toxicol. Jan;77(1):42-7.
- 16.Wang CJ, Wang JM, Lin WL, *et al* (2000): Protective effect of Hibiscus anthocyanins against tert-butyl hydroperoxide-induced hepatic toxicity in rats. <u>Food Chem Toxicol.</u> May;38(5):411-6.
- 17.Chewonarin T, Kinouchi T, Kataoka K, *et al* (1999): Effects of roselle (Hibiscus sabdariffa Linn.), a Thai medicinal plant, on the mutagenicity of various known mutagens in Salmonella typhimurium and on formation of aberrant crypt foci induced by the colon carcinogens azoxymethane and 2-amino-1-methyl-6-phenylimidazo[4,5-b]pyridine in F344 rats. Food Chem Toxicol. Jun;37(6):591-601.
- 18. Tseng TH, Kao TW, Chu CY, *et al* (2000): Induction of apoptosis by hibiscus protocatechuic acid in human leukemia cells via reduction of retinoblastoma (RB) phosphorylation and Bcl-2 expression. <u>Biochem Pharmacol</u>. Aug 1; 60(3):307-15.
- 19. Tseng TH, Hsu JD, Lo MH, *et al* (1998): Inhibitory effect of Hibiscus protocatechuic acid on tumor promotion in mouse skin. <u>Cancer Lett.</u> Apr 24;126(2):199-207.
- 20.Simon, J.E., Chadwick A.F. Craker L.E.. (1984): Herbs: An Indexed Bibliography. 1971-1980. The Scientific Literature on Selected Herbs, and Aromatic and Medicinal Plants of the Temperate Zone. Archon Books, 770 pp., Hamden, CT.
- 21. Iacobellis NS, Lo Cantore P, Capasso F, Senatore F. (2005): Antibacterial activity of Cuminum cyminum L. and Carum carvi L. essential oils. J Agric Food Chem. Jan 12;53(1):57-61.
- 22.Singh G, Kapoor IP, Pandey SK, *et al* (2002): Studies on essential oils: part 10; antibacterial activity of volatile oils of some spices. <u>Phytother Res.</u> Nov;16(7):680-2.
- 23.Harborne, J.B. (1973): Phytochemical methods . Science paperbacks, Chapman and Hall. London, UK.
- 24. Trease, GE.; Evans, WC. (1983): Pharmacognosy. London, Bailliere Tindall Press.. p. 309–706.
- 25.Harborn J.B. (1984): Phytochemical methods. Aguide to modern techniques of plant analysis. Chapman and Hall. London, UK.
- 26.Adeday O., Andewtson W., Moo-Young M., *et al* (2001): Phytochemistry and antibacterial activity of *Senna alata* flower. <u>Pharmaceutical Biology</u> 39:1-5).
- 27.Perez, C.; Pauli, M. and Bazerque, P. (1990): An antibiotic assay by the agar-well diffusion method. J.Actabiologiae, 15:113-115.
- 28.Bandyopadhyay D, Chatterjee TK, Dasgupta A, *et al* (2005): In vitro and in vivo antimicrobial action of tea: the commonest beverage of Asia. <u>Biol Pharm Bull.</u> 2005 Nov;28(11):2125-7.
- 29.Singh S, Malhotra M, Majumdar DK. (2005): Antibacterial activity of Ocimum sanctum L. fixed oil: Indian J Exp Biol. Sep;43(9):835-7.
- 30.Thuille N, Fille M, Nagl M. (2003): Bactericidal activity of herbal extracts. Int J Hyg Environ Health. Jun;206(3):217-21.
- 31.Juliani HR Jr, Biurrun F, Koroch AR, *et al* (2002): Chemical constituents and antimicrobial activity of the essential oil of Lantana xenica. <u>Planta Med.</u> Aug; 68(8):762-4.

- 32.Sonboli A, Salehi P, Kanani MR, Ebrahimi SN. (2005): Antibacterial and antioxidant activity and essential oil composition of Grammosciadium scabridum Boiss. from Iran <u>Z</u> <u>Naturforsch [C]</u>. Jul-Aug;60(7-8):534-8.
- 33.Vilasinee Hirunpanich¹⁾, Anocha Utaipat¹⁾, Noppawan Phumala Morales²⁾, Nuntavan *et al* (2005): Antioxidant Effects of Aqueous Extracts from Dried Calyx of *Hibiscus sabdariffa* LINN. (Roselle) *in Vitro* Using Rat Low-Density Lipoprotein (LDL) <u>Biological & Pharmaceutical Bulletin</u> Vol. 28, No. 3 481
- 34.Liu,-C-L; Wang,-J-M; Chu,-C-Y; Cheng,-M-T; Tseng,-T-H (2002): In vivo protective effect of protocatechuic acid on tert-butyl hydroperoxide-induced rat hepatotoxicity. <u>Food-Chem-Toxicol</u>. May; 40(5): 635-41.
- 35.Chanida HANSAWASDI¹⁾, Jun KAWABATA¹⁾ and Takanori KASAI¹ (2000): α-Amylase Inhibitors from Roselle (*Hibiscus <u>sabdariffa</u> Linn.*) Tea. <u>Bioscience</u>, <u>Biotechnology</u>, and <u>Biochemistry</u>. Vol. 64, No. 5 pp.1041-1043
- 36.Jonadet M, Bastide J, Bastide P, *et al* (1990): [In vitro enzyme inhibitory and in vivo cardioprotective activities of hibiscus (Hibiscus sabdariffa L.)] <u>J Pharm Belg.</u> Mar-Apr; 45(2):120-4. (English abstract)
- 37.Olukemi O. Ilori, Olukemi A. Odukoya (2005): *Hibiscus sabdariffa* and Sorghum bicolor as natural colorants. <u>Electronic J. Envir. Agric. Food Chemistry.</u> Indexed in CAS-Scifinder.
- 38.Haji Faraji M., Haji-Tarkhani A. J. Ethnopharmacol. 65,231–236 (1999)
- 39. Thomas Frank, Marlies Janßen, Michael Netzel, *et al* (2005): Pharmacokinetics of Anthocyanidin-3-Glycosides Following Consumption of *Hibiscus sabdariffa* L. Extract *Journal of Clinical Pharmacology*, ; 45:203-210.
- 40.Francis. F. J. (2000): <u>Cereals Foods World</u>. 45, 208–213
- 41.Kuroda,-K; Nishimura,-N; Izumi,-A; Dimmel,-D-R (2002): Pyrolysis of lignin in the presence of tetramethylammonium hydroxide: a convenient method for S/G ratio determination. J-Agric-Food-Chem. Feb 27; 50(5): 1022-7
- 42.Dafallah AA, Al-Mustafa Z (1996): Investigation of the anti-inflammatory activity of Acacia nilotica and Hibiscus sabdariffa. <u>Am J Chin Med</u>. ;24(3-4):263-9.
- 43.Brunold C, Deters A, Knoepfel-Sidler F, *et al* (2004): Polysaccharides from Hibiscus sabdariffa flowers stimulate proliferation and differentiation of human keratinocytes. <u>Planta Med.</u> Apr; 70(4):370-3.
- 44.Muller BM, Franz G. (1992): Chemical structure and biological activity of polysaccharides from Hibiscus sabdariffa. <u>Planta Med.</u> Feb;58(1):60-7.
- 45.Wrobel K, Wrobel K, Urbina EM. (2000): Determination of total aluminum, chromium, copper, iron, manganese, and nickel and their fractions leached to the infusions of black tea, green tea, Hibiscus sabdariffa, and Ilex paraguariensis (mate) by ETA-AAS. <u>Biol</u> <u>Trace Elem Res</u>. Winter;78(1-3):271-80.
- 46.Yan JH, Tang KW, Zhong M, Deng NH. (2002): [Determination of chemical components of volatile oil from Cuminum cyminum L. by gas chromatography-mass spectrometry]. <u>Se</u> <u>Pu.</u> Nov; 20(6):569-72. (English abstract)
- 47.Toru Ishikawa, Tomomi Takayanagi, Junichi Kitajima (2002): Water-Soluble Constituents of Cumin: Monoterpenoid Glucosides. Chem. Pharm. Bull. 50 (11) 1471.
- 48.Al-Bataina BA, Maslat AO, Al-Kofahil MM. (2003): Element analysis and biological studies on ten oriental spices using XRF and Ames test. J Trace Elem Med Biol.;17(2):85-90.
- 49.Nostro A, Cellini L, Di Bartolomeo S, Di Campli E, *et al* (2005): Antibacterial effect of plant extracts against Helicobacter pylori. <u>Phytother Res.</u> Mar;19(3):198-202.