



Histological changes of extract cumin cyminum on the leydig cells, seminiferous tubules and epididymis in adult rabbits (*Oryctolagus cuniculus*)

W.F. Obead¹ , H.H. Khalaf²  and H.B. Mahmood¹ 

¹Department of Anatomy, College of Veterinary Medicine, University of Kerbala, Kerbala, ²Department of Anatomy, College of Veterinary Medicine, University of Fallujah, Fallujah, Iraq

Article information

Article history:

Received 22 February, 2023
Accepted 20 November, 2023
Available online 01 January, 2024

Keywords:

Testis
Spermatogonia
Spermatid
Interstitial cell

Correspondence:

W.F. Obead
walaa.obead@uokerbala.edu.iq

Abstract

The primary histological alterations caused by using the cumin plant and associated with rabbit fertilization are the subject of the present investigation. The current study used ten mature, healthy male rabbits have been readied for testing, with five rabbits in each treatment and control group. The treatment group received an extract from the cumin plant at a dose of 250 mg/kg b.w/orally administered once daily for six weeks, while the control group received a regular diet and water. An extract was made from each 25 g of dried cumin powder, and it was subsequently diluted with 10 ml of boiling distilled water. The extraction process took thirty minutes to be finished. After filtering each extract, its concentration was adjusted to provide 250 mg/ml of sample per milliliter. After that, the solutions are kept in a container that is tightly closed. The primary histological changes detected by hematoxylin and eosin in this study were damage to seminiferous tubules and a decrease in the thickness of the germinal layer of spermatogonia. In addition, the Leydig cells (Interstitial cells) in the interstitial space were weak and deteriorating. Moreover, the arrangement of epithelial cells in the epididymis duct with empty spermatozoa was seen. Cumin consumption in large doses causes a lot of histological changes that show up in the male reproductive system.

DOI: [10.33899/ijvs.2023.138536.2811](https://doi.org/10.33899/ijvs.2023.138536.2811), ©Authors, 2023, College of Veterinary Medicine, University of Mosul.
This is an open access article under the CC BY 4.0 license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

Herbs and other components can treat or ward off illness, as is well-known (1-3). Due to their accessibility, safety, and use, some edible herbal plant species, including cumin (*Cuminum cyminum* L.), are frequently employed as food additives. Cumin contains two numbers (14 chromosomes) and is a component of the flowering plant, tribe Ammineae, and subtribe Carinae (4-6). After black pepper, cumin is the second most commonly used seed kind. It is an annual plant with several therapeutic, nutraceutical, and pharmacological characteristics. It is among the oldest and most widely used aromatic and herbaceous natural products. Additionally, cumin is commonly used in the culinary, beverage, alcohol, pharmaceutical, cosmetic, and toiletry sectors (7,8). Despite

the common uses of cumin in many medical fields as anti-diabetic and antimicrobial activity, its frequent use damages the male reproductive system. It leads to histological changes (9). Some inflammatory markers that treatments supplemented with *C. cyminum* have been demonstrated to impact significantly are adiponectin, high sensitivity C-reactive protein (hs-CRP), and TNF (10-12). This article extensively discusses *C. cyminum* generally beneficial properties (13). The use of ingredients to flavor various food compositions, including cheese, pickles, soups, beans, and alcoholic drinks, is widespread. Cumin is an aromatic herb commonly used as a food supplement globally. It is also employed in herbal medicine, particularly veterinary medicine (14-16). Seeds have demonstrated antioxidant qualities and help treat colic, diarrhea, and cholera (17).

Cumin seed extract is also a microorganism-fighting oil (18,19). Most medicines and other active pharmacological compounds now under development originate from the natural kingdom. Given the high toxicity of various synthetic medications, the therapeutic use of herbal medicines has increased over the recent decade, dramatically increasing the number of herbal medicine makers (20,21). The cumin compound has been shown to have active antioxidant properties, which can restrain lipid peroxides and free radicals (22). In diabetic rats, it possesses the capacity to lower plasma lipid profile (23).

Even though cumin is often used in medicine, too much might lead to hormonal abnormalities; therefore, this study was done to recognize the most significant histological changes.

Materials and methods

Experimental design

In the current study, ten adult male local rabbits weighing approximately 1150-1300 g have always been separated into two groups (5 group). The control group received a typical diet and water, whereas the treatment group received cumin extract 250 mg/kg b.w. through stomach tube feeding (24). For all groups, the experiment lasted for six weeks. By administering intramuscular injections of xylazine 0.5 ml and ketamine, all study rabbits were killed 0.5 ml.

Cumin extraction

Cumin powders were purchased from the Kerbala city market. Each 25 g of dry cumin powder was used to prepare an extract, after which it was treated with 10 ml of boiling distilled water. The extraction process required thirty minutes. Each extract had been filtered, and the content was modified so that each ml of sample was equal to 250 mg/ml. Those solutions are subsequently stored in a tightly sealed container (25).

Histological preparations

The testis served as the source of the anatomical samples. The samples, which were about 0.5 cm in size, were then preserved in 10% Neutral buffered formalin for 48 hours. Hematoxylin and eosin stain were used with conventional histological methods to distinguish distinct tissue components (26). The germinal layer of the seminiferous tubules and the interstitial space occupied by Leydig cells and all tissue sections have been incorporated into the histological measurements. The histological characteristics were analyzed with the help of a digital USB microscopic camera (Canon 550D, 18 Megapixel, Japan), which was attached to a Novel microscope. A stage micrometer has been employed to correlate the objective lenses with the program (27).

Ethical approve

Under the reference number UOK.VET.AN.2022.052, this research was carried out in the anatomical laboratory of the College of Veterinary Medicine at the University of Kerbala.

Results

Histological features

The illustration shows the typical characteristics of the testes in the control group, including regular histological features with well-developed spermatids and a germ cell lining performing cell proliferation. Furthermore, we observed Sertoli cell activity and their presence in their usual location in the basement membrane of seminiferous tubules, which revealed the arrangement of spermatogonial stages. The mean diameters of seminiferous tubules were approximately 98.14 μm (Figure 1). In the control group, Leydig cells were observed to be active; these cells appeared as a massive regular, with non-granular and spherical cytoplasm and abundant nuclei with visible nucleoli. The majority of the nuclei are located in the cytoplasm's middle. The mean thickness of the Leydig area was 152.21 μm (Figure 2). Moreover, the histological features of the rabbit epididymis duct in the control group were lined by pseudostratified columnar epithelium and a large number of spermatozoa in the epididymis lumen (Figure 3).

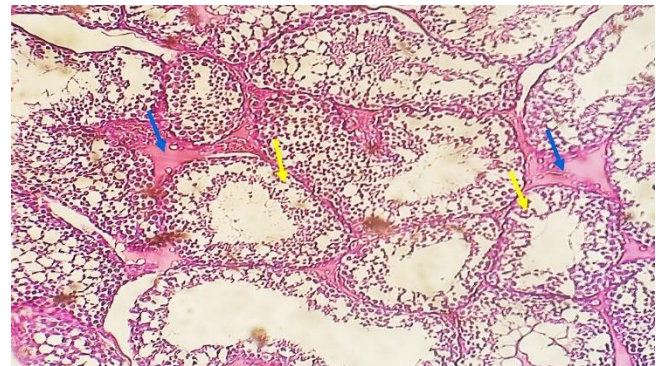


Figure 1: Photomicrograph of control testis in rabbits showing the normal histological features in seminiferous tubules (yellow arrows), and activity secretion in interstitial space (blue arrows). H&E stain 10x.

When the histological testis characteristics of the extract-cumin group were examined under the microscope, numerous changes were observed, including the destruction of the seminiferous tubules, a significant number of damaged cells, the absence of spermatozoa in the tubule lumen output, spermatogonia detaching from the basal membrane in some areas, and an increase in the space between the seminiferous tubules, the mean diameters of seminiferous tubule about 50.22 μm (Figure 4). In this Group, we observed the decrease

in the number of Leydig cells an appearance of inactivity, limited cytoplasm, and psychotic nucleus the mean thickness of Leydig area was 92.45 μm (Figure 5). The epididymis duct appears empty in this group due to spermatozoa in the epididymis lumen (Figure 6).

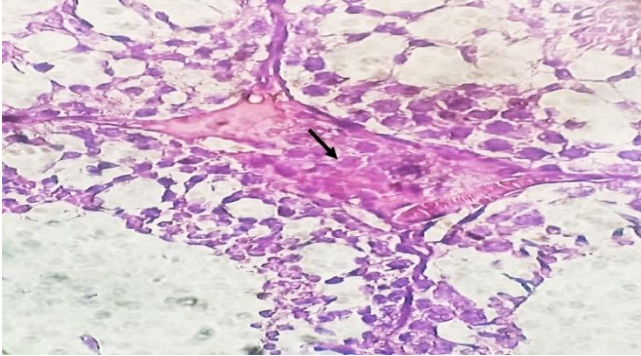


Figure 2: Photomicrograph of control testis in rabbits showing the crowded and activity in leydig cells (black arrow). H&E stain 40x.

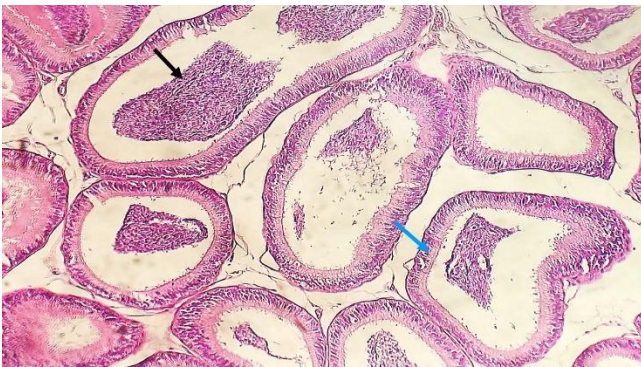


Figure 3: Photomicrograph of control epididymis in rabbits showing a lot of sperms in ducts of epididymis (black arrow), and activity lining epithelia (blue arrow). H&E stain 10x.

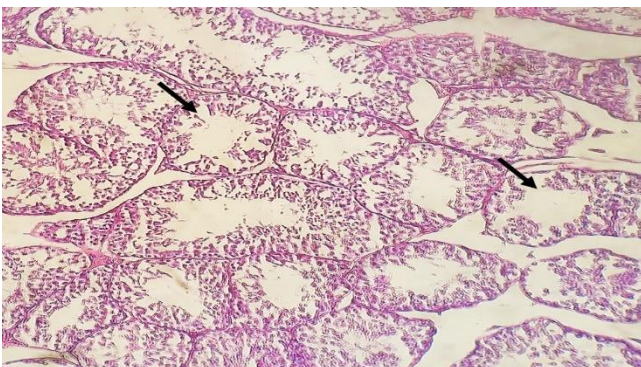


Figure 4: Photomicrograph of extract cumin testis in rabbits showing the damage in semiferioius tubules (black arrows). H&E stain 10x.

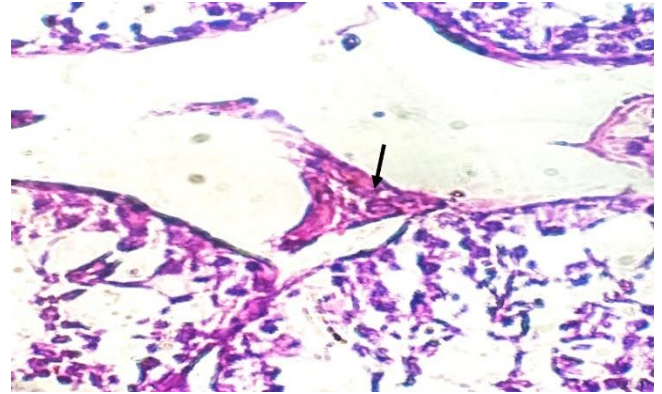


Figure 5: Photomicrograph of extract cumin testis in rabbits showing the leydig cells in active and smaller area size (black arrow). H&E stain 40x.



Figure 6: Photomicrograph of extract cumin epididymis in rabbits showing of ducts of epididymis from sperms (black arrow), and inactivity lining epithelia (blue arrow). H&E stain 10x.

Discussion

The current study shows the typical appearances of the testis in the control group, including regular histological features with very well-developed germ cell lining performing cell proliferation. In contrast, the current study found numerous modifications in the extract-cumin group, including destruction to the seminiferous tubules, a significant number of damaged cells, the absence of spermatozoa in the tubule lumen output, spermatogonia detaching from the basal membrane in some areas, and an increase in the space between the seminiferous tubules. This outcome is similar to that of Abdi *et al.* (28) and Taha (29), which mentions that in a healthy condition, spermatogonia and the basement membrane are nearby in the seminiferous tubules, where germ cells are alive and appear active. Unnoticed changes also included the degradation of specific spermatogenic cells and an accumulation in the tubular

hollow. These significances might have resulted from excessive consumption of cumin extract, which is widely used as a body slimming agent and may have caused weight loss.

The control group observed regular histological features with the activity of Sertoli cells in the control group; additionally, the Leydig cells had nuclei located in the cytoplasm's middle, a wide interstitial area that was occupied by Leydig cells, whereas cumin describes a decrease in the number of Leydig cells, inactivity appearance, limited cytoplasm, and psychotic nuclei in the extract group. Findings are akin to Tripepi *et al.* (30) and Malhi *et al.* (31) those that describe the Leydig cells. They were found in groups or cords of various shapes and sizes with round nuclei and granular cytoplasm. These cells seem to be distributed randomly, some being perivascular, others related to vessels. The testicular hormone was markedly lower in these results compared to the control group, comparable to those of Willatgamuwa *et al.* (32), which evaluated the therapy with a water extract of the cumin plant. These studies, however, contradict Smith *et al.* (33), Hipler *et al.* (34) and Fonseca *et al.* (35) that claimed that the Cumin therapy was ineffective when administered alone in sperm characteristics and had a detrimental impact. Gupta *et al.* (36), Sharifi *et al.* (37) and Al-Hafedh *et al.* (38). The testicular tissue may have acquired changes due to all these histological changes caused by hormone changes. Consequently, these variations need the cells responsible for adapting these mechanisms to maintain the new metabolic balance.

The present study explains why the epididymis duct in the extract cumin team appeared to be vacant of spermatozoa, in contrast to the morphological features of the rabbit epididymis duct in the comparison group, which were lined by a significant number of spermatozoa in the epididymis lumen and had a regular appearance of pseudostratified columnar epithelium. Our findings are consistent with those of others who believed that some variables determined by sperm concentration are elements of sperm parameters that significantly affect male fertility Roshankhah *et al.* (39), Al-Allaf *et al.* (40) and Amin *et al.* (41). Herbal medicine may be widely used in many cultures to treat conditions like obesity and to enhance sperm quality. Hence, excessive use always has a harmful impact on the amount and quality of sperm production.

Conclusion

It is clear that eating unbearable cumin in large amounts results in a variety of histological alterations in the male reproductive system. The study concluded that cumin extract 250 g/kg b.w. caused germ cell damage in seminiferous tubules and Sertoli and Leydig cell degeneration, resulting in rabbit infertility.

Acknowledgment

The College of Veterinary Medicine at the University of Kerbala is acknowledged for facilitating this investigation.

Conflict of interest

There is absolutely no apparent conflict, according to the author.

References

1. Jafari S, Sattari R, Ghavamzadeh SI. Evaluation the effect of 50 and 100 mg doses of *Cuminum cyminum* essential oil on glycemic indices, insulin resistance and serum inflammatory factors on patients with diabetes type II: A double-blind randomized placebo-controlled clinical trial. *J Tradit Complement Med.* 2017;7(3):332-338. DOI: [10.1016/j.jtcme.2016.08.004](https://doi.org/10.1016/j.jtcme.2016.08.004)
2. Ahmed IA, Mikail MA, Bin Ibrahim M, Bin Hazali N, Rasad MS, Ghani RA, Wahab RA, Arief SJ, Yahya MN. Antioxidant activity and phenolic profile of various morphological parts of underutilised *Baccaurea angulata* fruit. *Food Chem.* 2015;172(1):778-787. DOI: [10.1016/j.foodchem.2014.09.122](https://doi.org/10.1016/j.foodchem.2014.09.122)
3. Ibrahim M, Ahmed IA, Mikail MA, Ishola AA, Draman S, Isa ML, Yusuf AM. *Baccaurea angulata* fruit juice reduces atherosclerotic lesions in diet-induced Hypercholesterolemic rabbits. *Lipids Health Dis.* 2017;16:1-8. DOI: [10.1016/j.foodchem.2014.09.122](https://doi.org/10.1016/j.foodchem.2014.09.122)
4. Allaq AA, Sidik NJ, Abdul-Aziz A, Ahmed IA. Cumin (*Cuminum cyminum* L.): A review of its ethnopharmacology, phytochemistry. *Biomed Res Ther.* 2020;7(9):4016-4021. DOI: [10.15419/bmrat.v7i9.634](https://doi.org/10.15419/bmrat.v7i9.634)
5. Mikail MA, Zamakshshari N, Abdullah AH. Natunti-aging skincare: Role and potential. *Biogerontol.* 2020;21:293-310. DOI: [10.1007/s10522-020-09865-z](https://doi.org/10.1007/s10522-020-09865-z)
6. Rebey IB, Bourgou S, Rahali FZ, Msaada K, Ksouri R, Marzouk B. Relation between salt tolerance and biochemical changes in cumin (*Cuminum cyminum* L.) seeds. *J Food Drug Anal.* 2017;25(2):391-402. DOI: doi.org/10.1016/j.jfda.2016
7. Bhatt J, Kumar S, Patel S, Solanki R. Sequence-related amplified polymorphism (SRAP) markers based genetic diversity analysis of cumin genotypes. *Ann Agrar Sci.* 2017;15(4):434-438. DOI: [10.1016/j.aasci.2017.09.001](https://doi.org/10.1016/j.aasci.2017.09.001)
8. Thippeswamy N, Naidu KA. Antioxidant potency of cumin varieties-cumin, black cumin and bitter cumin-on antioxidant systems. *Eur Food Res Technol.* 2005;220(56):472-476. DOI: [10.1007/s00217-004-1087-y](https://doi.org/10.1007/s00217-004-1087-y)
9. El-Said AH, Goder EH. Antifungal activities of *Cuminum cyminum* and *Pimpinella anisum* essential oils. *Int J Curr Microbiol Appl Sci.* 2014;3(3):937-44. DOI: [10.1016/S0308-8146\(01\)00326-0](https://doi.org/10.1016/S0308-8146(01)00326-0)
10. Noori H, Moosavi SG, Seghatoleslami M, Rostampour MF. Responses of cumin (*Cuminum cyminum* L.) to different seed priming methods under osmotic stress. *Not Bot Horti Agrobot Cluj Napoca.* 2022;50(1):12600. DOI: [10.15835/nbha50112600](https://doi.org/10.15835/nbha50112600)
11. Soltani E, Mortazavian SM, Faghghi S, Akbari GA. Non-deep simple morphophysiological dormancy in seeds of *Cuminum cyminum* L. *J Appl Res Med Aromat Plants.* 2019;15:100222. DOI: [10.1016/j.jarmap.2019.100222](https://doi.org/10.1016/j.jarmap.2019.100222)
12. Kalaiyani P, Saranya RB, Ramakrishnan G, Ranju V, Sathiya S, Gayathri V, Thiyagarajan LK, Venkatesh JR, Babu CS, Thanikachalam S. *Cuminum cyminum*, a dietary spice, attenuates hypertension via endothelial nitric oxide synthase and NO pathway in renovascular hypertensive rats. *Clin Exp Hypertens.* 2013;35(7):534-42. DOI: [10.3109/10641963.2013.764887](https://doi.org/10.3109/10641963.2013.764887)
13. Srinivasan K. Cumin (*Cuminum cyminum*) and black cumin (*Nigella sativa*) seeds: Traditional uses, chemical constituents, and nutraceutical effects. *Food Qual Saf.* 2018;2(1):1-16. DOI: [10.1093/fqsafe](https://doi.org/10.1093/fqsafe)

14. Sarfraz RA, Hassan QU, Murtaza S. Effect of different solvents on the chemical composition and anti-diabetic activity of *Acacia arabica* and *Zizyphus mauritiana*. J Med Herbs Ethnomed. 2018;(4):35-37. DOI: [10.25081/jmhe.2018.v4.3638](https://doi.org/10.25081/jmhe.2018.v4.3638)
15. Lai PK, Roy J. Antimicrobial and chemopreventive properties of herbs and spices. Curr Med Chem. 2004;11(11):1451-1460. DOI: [10.2174/0929867043365107](https://doi.org/10.2174/0929867043365107)
16. Zheng J, Zhou Y, Li Y, Xu DP, Li S, Li HB. Spices for prevention and treatment of cancers. Nutrients. 2016;12(8):495. DOI: [10.3390/nu8080495](https://doi.org/10.3390/nu8080495)
17. El-Ghorab AH, Nauman M, Anjum FM, Hussain S, Nadeem M. A comparative study on chemical composition and antioxidant activity of ginger (*Zingiber officinale*) and cumin (*Cuminum cyminum*). J Agric Food Chem. 2010;58(14):8231-8237. DOI: [10.1021/jf101202x](https://doi.org/10.1021/jf101202x)
18. Daljit SA, Jasleen K. Antimicrobial activity of spices. Int J Antimicrob Agents. 1999; 12(3):257-262. DOI: [10.1016/S0924-8579\(99\)00074-6](https://doi.org/10.1016/S0924-8579(99)00074-6)
19. Abbaszadegan A, Gholami A, Ghahramani Y, Ghareghan R, Ghareghan M, Kazemi A, Iraj A, Ghasemi Y. Antimicrobial and cytotoxic activity of *Cuminum cyminum* as an intracanal medicament compared to chlorhexidine gel. Iran Endod J. 2016;11(1):44. DOI: [10.7508/iej.2016.01.009](https://doi.org/10.7508/iej.2016.01.009)
20. Kumari R, Kotecha M. A review on the Standardization of herbal medicines. Int J Pharma Sci Res. 2016;7(2):97-106. [\[available at\]](#)
21. Nasreen S, Radha R. Assessment of quality of *Withania somnifera* Dunal (Solanaceae) Pharmacognostical and physicochemical profile. Int J Pharm Pharm Sci. 2011;3(2):152. DOI: [10.22159/ijpps.2018v10i2.22915](https://doi.org/10.22159/ijpps.2018v10i2.22915)
22. Rice-Evans CA, Diplock AT. Current status of antioxidant therapy. Free Radic Biol Med. 1993;15(1):77-96. DOI: [10.1016/0891-5849\(93\)90127-G](https://doi.org/10.1016/0891-5849(93)90127-G)
23. Dhandapani S, Subramanian VR, Rajagopal S, Namasivayam N. Hypolipidemic effect of *Cuminum cyminum* L. on alloxan-induced diabetic rats. Pharmacol Res. 2002;46(3):251-255. DOI: [10.1016/S1043-6618\(02\)00131-7](https://doi.org/10.1016/S1043-6618(02)00131-7)
24. hmed Aljanabi MS, Alfahdawi OA, Ismail TF. Physiological and histological study of the effect of Cumin Cyminum watery extract and vitamin E on the male reproductive system in rats exposed to oxidative stress. Tikrit J Pure Sci. 2018;23(3):23-32. DOI: [10.25130/tjps.23.2018.044](https://doi.org/10.25130/tjps.23.2018.044)
25. Al-Saaedi AM. Antipyretic activity of the aqueous extract of Cumin (*Cuminum cyminum* L.) with yeast induced pyrexia in female rats. Univ Thi-Qar J Sci. 2021;8(1):33-5. DOI: [10.32792/utq/utjs/vol8/1/5](https://doi.org/10.32792/utq/utjs/vol8/1/5)
26. Obead WF, Dawood GA, Mahmood HB. A Comparative histological study of the soft palate between rabbits and guinea pigs. HIV Nurs. 2022;22(2):1075-1077. [\[available at\]](#)
27. Sultan GA, Al-Haaik AG, Alhasso AA. Morphometrical and histochemical study of glandular stomach (Proventriculus) in local domestic male ducks (*Anas platyrhynchos*). Iraqi J Vet Sci. 2023;37(1):65-71. DOI: [10.33899/ijvs.2022.133451.2233](https://doi.org/10.33899/ijvs.2022.133451.2233)
28. Abdi OA, Glover EK, Luukkanen O. Causes and impacts of land degradation and desertification: Case study of the Sudan. Int J Agric For. 2013;3(2):40-51. DOI: [10.5923/j.ijaf.20150506.04](https://doi.org/10.5923/j.ijaf.20150506.04)
29. Taha AN, Ismail HK. The impact of nano zinc oxide particles on the histology of the male reproductive system of adult male rabbits. Iraqi J Vet Sci. 2023; 37(1):105-113. DOI: [10.33899/ijvs.2022.133632.2270](https://doi.org/10.33899/ijvs.2022.133632.2270)
30. Tripepi S, Carelli A, Perrotta E, Brunelli E, Tavolario R, Facciolo RM, Canonaco M. Morphological and functional variations of Leydig cells in testis of the domestic pig during the different biological stages of development. J Exp Zool. 2000;287(2):167-75. DOI: [10.1002/1097-010X\(20000701\)287:2<167::AID-JEZ7>3.0.CO;2-3](https://doi.org/10.1002/1097-010X(20000701)287:2<167::AID-JEZ7>3.0.CO;2-3)
31. Malhi PS, Roy KS, Pawar HS. Histomorphological studies on the postnatal development of seminiferous tubules in testis of Indian Murrah buffalo (*Bubalus bubalis*). Buffalo J. 1999;15:215-224. DOI: [10.1016/j.theriogenology.2009.06.034](https://doi.org/10.1016/j.theriogenology.2009.06.034)
32. Willatgamuwa SA, Platel K, Saraswathi G, Srinivasan K. Antidiabetic influence of dietary cumin seeds (*Cuminum cyminum*) in streptozotocin induced diabetic rats. Nutr Res. 1998;18(1):131-42. DOI: [10.1016/S0271-5317\(97\)00207-8](https://doi.org/10.1016/S0271-5317(97)00207-8)
33. Smith SJ, Lopresti AL, Teo SY, Fairchild TJ. Examining the effects of herbs on testosterone concentrations in men: A systematic review. Adv Nutr. 2021;12(3):744-765. DOI: [10.1093/advances/nmaa134](https://doi.org/10.1093/advances/nmaa134)
34. Hipler UC, Görnig M, Hipler B, Römer W, Schreiber G. Stimulation and scavestrogen-induced inhibition of reactive oxygen species generated by rat Sertoli cells. Arch Androl. 2000;44(2):147-154. DOI: [10.1080/014850100262326](https://doi.org/10.1080/014850100262326)
35. Fonseca VA, Stone A, Munshi M, Baliga BS, Aljada A, Thusu K, Fink L, Dandona P. Oxidative stress in diabetic macrovascular disease: does homocysteine play a role. Southern medical journal. 1997; 90 (9): 903-906. DOI: [10.1097/00007611-199709000-00008](https://doi.org/10.1097/00007611-199709000-00008)
36. Gupta RS, Saxena P, Gupta R, Kachhawa JB. Evaluation of reversible contraceptive activities of *Cuminum cyminum* in male albino rats. Contraception. 2011;84(1):98-107. DOI: [10.1016/j.contraception.2010.10.013](https://doi.org/10.1016/j.contraception.2010.10.013)
37. Sharifi-Rad J, Rayess YE, Rizk AA, Sadaka C, Zgheib R, Zam W, Sestito S, Rapposelli S, Neffe-Skocińska K, Zielińska D, Salehi B. Turmeric and its major compound curcumin on health: Bioactive effects and safety profiles for food, pharmaceutical, biotechnological and medicinal applications. Front Pharmacol. 2020;11:01021. DOI: [10.3389/fphar.2020.01021](https://doi.org/10.3389/fphar.2020.01021)
38. Al-Hafedh SO, Cedden F. The impact of various antioxidant supplementation on ram's sperm quality, fertilization, and early embryo development, in vitro. Iraqi J Vet Sci. 2022;36(4):869-76. DOI: [10.33899/ijvs.2022.132426.2092](https://doi.org/10.33899/ijvs.2022.132426.2092)
39. Roshankhah SH, Salahshoor MR, Aryanfar S, Jalili F, Sohrabi M, Jalili C. Effects of curcumin on sperm parameters abnormalities induced by morphine in rat. J Med Biomed Sci. 2017;6:1-10. DOI: [10.4314/jmbs.v6i2.1](https://doi.org/10.4314/jmbs.v6i2.1)
40. Al-Allaf LI, Sultan ON, Saad-Allah BS, Al-Nuaimy WM. Does Baclofen induce changes in testicular histology and seminal fluid analysis in rat?. Iraqi J Vet Sci. 2021;35(2):387-96. DOI: [10.33899/ijvs.2020.126894.1411](https://doi.org/10.33899/ijvs.2020.126894.1411)
41. Amin KA, Nagy MA. Effect of Carnitine and herbal mixture extract on obesity induced by high fat diet in rats. Diabetol Metab Syndr. 2009;1(1):1-4. DOI: [10.1186/1758-5996-1-17](https://doi.org/10.1186/1758-5996-1-17)

التغيرات النسيجية لمستخلص الكمون على خلايا لايدك، النبيبات المنوية والبربخ في الارانب البالغة

ولاء فاضل عبيد^١، حارث حمادي خلف^٢ و حسين بشار محمود^١

^١ فرع التشريح، كلية الطب البيطري، جامعة كربلاء، كربلاء، أفرع
التشريح، كلية الطب البيطري، جامعة الفلوجة، الفلوجة، العراق

الخلاصة

إن التغيرات النسيجية الأولية الناتجة عن استخدام نبات الكمون والمرتبطة بالإخصاب في الأرانب هي موضوع التحقيق الحالي. أخذت في الدراسة الحالية عشرة أرانب ذكور سليمة ناضجة للاختبار، خمسة أرانب لكل من مجموعة العلاج والسيطرة. تلقت مجموعة العلاج مستخلص نبات الكمون بجرعة ٢٥٠ مجم / كجم من وزن الجسم / عن طريق الفم مرة واحدة يومياً لمدة ٦ أسابيع بينما تلقت المجموعة السيطرة حمية طبيعية وماء. تم تحضير المستخلص ٢٥ غم من مسحوق الكمون المجفف، وتم تخفيفه بعد ذلك بـ ١٠ مل من الماء المقطر المغلي. استغرقت عملية الاستخلاص نحو ثلاثون دقيقة بعد ذلك تم تصفيته للتخلص من الشوائب العالقة، تم ضبط تركيزه لتجهيز ٢٥٠ ملغم/مل من المحلول المراد تجريبه. بعدها، تم حفظ المحلول في حاوية مغلقة بإحكام. أهم التغيرات النسيجية الأولية التي تم الكشف عنها كانت بواسطة استخدام

ومنتكسة. علاوة على ذلك، كان ترتيب للخلايا الظهارية غير منتظم وقناة البربخ الخالية من الحيوانات المنوية ومن الواضح أن استهلاك الكمون بجرعات كبيرة يسبب الكثير من التغيرات النسيجية التي تظهر في الجهاز التناسلي الذكري.

صبغة الهيماتوكسيلين والأبوسين بينت الدراسة الحالية كان هناك تلف كبير واضح جدا في نسيج الأنابيب المنوية وكذلك انخفاض واضح في سمك الطبقة الجرثومية للحيوانات المنوية. بالإضافة إلى ذلك، كانت خلايا لايديك (الخلايا البينية) في الفراغات النسيجية البينية ضعيفة