www.qu.edu.iq/journalvm



Research article

Haemonchus contortus: Review of recent molecular advances in anthelmintic resistance

and vaccination

Nuha Qasim Mohammed¹ Hiba Shehab Ahmed² Noor Idan Jarad¹ Amal Hassan Al-Shabbani³

- 1- College of Veterinary Medicine, University of AL-Qadisiyah, Iraq.
- 2- College of Biotechnology, University of AL-Qadisiyah, Iraq.
- 3- College of Pharmacy, University of AL-Qadisiyah, Iraq.

Corresponding Author Email: nuha.allban@qu.edu.iq

Abstract

Haemonchus contortus is one of the world's major financial worms that attack ruminants. It is a blood-sucking parasite founded in the abomasum, particularly in cattle, sheep, and goats. Nematode infections may cause anemia, weight loss, or even death in animals that are severely affected. Current management practices against H. contortus largely depend on regular anthelmintic therapies in all countries with varying incidence across various regions.

H. contortus thus aims to form new action techniques in order to overcome resistance to anthelmintic agents. One option is a logical approach focused on a thorough knowledge of the molecular pathways in growth and reproduction cycles in the manufacture in modern anti-parasite drugs and vaccines. Key molecules may be defined as potential drug targets by a simple description of molecular, biochemical functions.

Besides, it is immediately essential to formulate immunological control of farm animal nematode infections. Important prevention has been accomplished after vaccination with native protein extracts, which shows that vaccination is possible. This paper explores the success of H. contortus science in the world. Particular fields of concern include epidemiological research, genetic analysis, and anthelmintic resistance identification using traditional and molecular techniques; morphological and chemical research of crucial molecules in mechanism expansion, parasitic organism-host interaction, and vaccine research. In the suggested form of these opinions, areas for potential exploration and alternatives for new or revised prevention strategies are described.

Keywords: Control, Vaccination, genetic, *Haemonchus contortus*, Ruminant.

Introduction

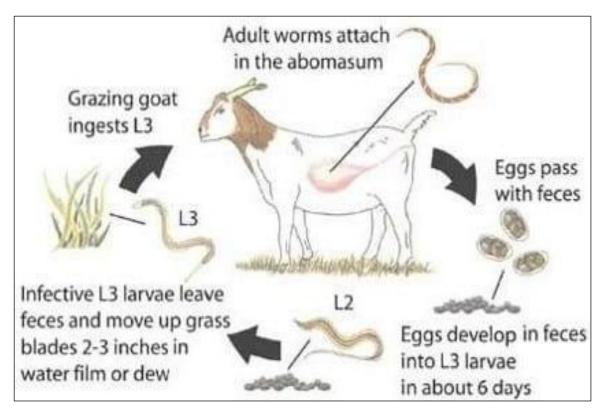
Nematodes have developed to take advantage of a wide variety of natural habitats. Although most stabilize a free lifestyle to reach maturity (Fig. 1), they depend on one or more hosts (1). A complex sequence of morphological features is taking place in several parasitic nematodes associated with migration from their hosts to create a mature infestation (2). Life cycles may include transitional hosts, vectors and, environmental-based stay when confronted with extreme and

unpredictable circumstances. including freezing or dehydration (3). The risks, namely predation, atmosphere, and immune reactions, of a wide variety, both plant and animal hosts have been adapted (1–3). The evolution of parasites affects public health (which amounts to 10 million life-years adjusting to disabilities), or indirectly by major financial failures in plant and animal production and by investing parasite in control (4).Anthelmintic medications are used as a

www.qu.edu.iq/journalvm



monitor, provided recurrently in livestock and through large-scale medical preventive initiatives (5). Although the effectiveness of these approaches was initially inevitable, the appearance or documenting inefficiency in human-infectious organisms of medicinal-resistant veterinary worms challenges continuous prevention efforts (4–7)



 $Fifure 1: The life cycle of {\it Haemonchus contortus}. {\it https://u.osu.edu/sheep/2019/07/30/ag-note-parasites-focus-haemonchus-contortus}.$

Vaccines provide an enticing alternative method for these parasites: control however, while two approved vaccines are presently accessible for veterinary purposes, the vast parasite heterogeneity and immune procedural mechanisms are significantly hindered in the production of vaccine; transcriptomic plasticity after the vaccine attempt may be used to prevent infections (8). It is thus obvious that modern, efficient control methods are essential. Helminths can respond to control measures and therefore evade them because of genetic variation (9). A deeper understanding of the nature and dynamics of this diversity across its spectrum can provide better knowledge of the pathways by which they are modified and can

recognize new goals that can be manipulated for influence (8,9).

The *H. contortus* (trichostrongylide) is a gastrointestinal worm of ruminants in tropical and temperate areas around the world, which has substantial effects on the economic and animal health of the sheep farm in a unique way (10). It also emerges as a prototype of a parasite nematode framework for functional and comparative genome sequencing, primarily due to its quick capability to collect resistance to medicines, its nearly equivalent tractability in controlled circumstances, the growth of enormous genomic inputs, and its special bond with other veterinary and medical nematodes of clad V (12,13).

www.qu.edu.iq/journalvm



Resistance to anthelmintics

The fundamental strategy for the containment of parasitic nematodes like H. contortus is focused upon the utilization of anthelmintics. but an anthelmintic resistance has over-grown. Resistance to anthelmintic in several areas of the planet was identified and published (6). A concise-short study of the nature of anthelmintic resistance in several nations refers to the occurrence of the resistance in countries like the United States, Mexico, South Africa, Australia, New Zealand, European countries. and China. Albendazole, fenbendazole, and ivermectin were used to treat in dairy goat farm (400 Goats) with confirmed infection (all above 80%) (14). Albendazole and ivermectin (IM) showed low activity both. fenbendazole displayed high effectiveness (6). This study showed a fecal egg count reduction test (FECRT) of 23,72% for the routine dose rate (5 mg /kg) albendazole. In this field, ivermectin has not been successful, with a prescribed FECR of 52.29 percent (0.2 mg/kg) and a 2-fold FECR of 89 percent (0.4 mg/kg) (6,14). The drug resistance was tested using albendazole, levamisole, ivermectin, a mixture of albendazole and ivermectin to check performance against nematode. The findings successful in the cases of levamisole, ivermectin, and the combination. The achievement of an FECR was above 95% and an FECR of albendazole of below 95% (6.14).

Molecular identification of anthelmintics resistance

Resistance to Benzimidazole

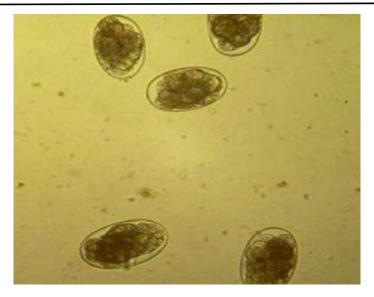
Three specific β -tubulin SNPs were seen to be linked to H. contortus with resistance to benzimidazole (BZ),

identified as F167Y (TTC to TAC), E198A (GAA to GCA) and F200Y (TTC to TAC) (15). A nucleotide sequence modification has resulted in alteration in the target protein and decreased BZ binding to the βtubulin (16). In five H. contortus species, including two sheep and three goats species, a multiplex PCR for the β-tubulin consistent with BZ resistance has been established, which suggests that the resistant genetic makeup is not found in any observed community of the nematode (15–20). In the meantime, an analysis utilizing PCR-SSCP in the area of the F200Y community H. contortus of sheep showed a plurality of homozygous prone genotype (21). Also, one analysis employed PCR-RFLP system to detect F200Y in H. contortus sheep populations, which revealed that the homozygous prone genotype is the most common (21). In various regions, a resistance was earlier revealed in the detection of all three known SNPs in a β-tubulin gene with PCRcoupled sequencing, which has been commonly used for the management of worm load (21).

Molecular approaches may be utilized, but they could not test resistance at the quantitative rates, as a diagnostic aid in detecting BZ resistance in populations (22). In fact, in combination with this field, molecular testing and biological testing are to be used (23). To date, the molecular research of the identification of BZ resistance in field samples, including several species of nematodes, has not been tested explicitly for this reason (22). Moreover, in fecal samples that included nematode eggs (Fig. 2) directly, molecular trials were not used, which could save a great deal of time. For now and on, whether current approaches or modern molecular production are used (22,23).

www.qu.edu.iq/journalvm





Fifure (2): The egg shape of Haemonchus contortus. <a href="https://www.researchgate.net/publication/262932569_Efficacy_and_Safety_of_Albendazole_against_Haemonchus_Contortus_Infestation_in_Goats/figures?lo=1&utm_source=google&utm_medium=organic

Resistanse to Ivermectin

the IM also relates to leading anthelmintic groups and has been used disproportionately, contributing widespread resistance (24). In addition to numerous genes that code for IM targets and efflux pumps, resistance to IM is considered to be very multi-gene based nature (25). SNPs around the entire genome were scanned using a 2b-RAD sequencing tool for further IM resistanceassociated SNPs in goats in both sensitive and resistant nematodes (24-26). 2962 SNPs and 2667 SNPs in resistant isolates have been identified (27). Among resistant or sensitive species, identical and relatively smaller genetic variations were observed (28). However, 208 SNPs with a significant variation, 24 of which were SNPs (29). This process is likely to be the primary selection of IM, with seven of the nines genetic markers expected to code for those proteins, which may play a crucial role in the IM target or efflux pump and even in receptor complex factor proteins, including membrane or neurons for transcriptional regulating proteins (30). It was suspected that these genes are correlated with IM tolerance. The findings

of this research revealed genes affecting the detection of IM and correlated with IM resistance in the genome of the nematode (31). Extensive SNP analysis utilizing the 2b-RAD sequencing method may be used to classify the worm (31).

Haemonchus contortus and diapausepresented genetic markers

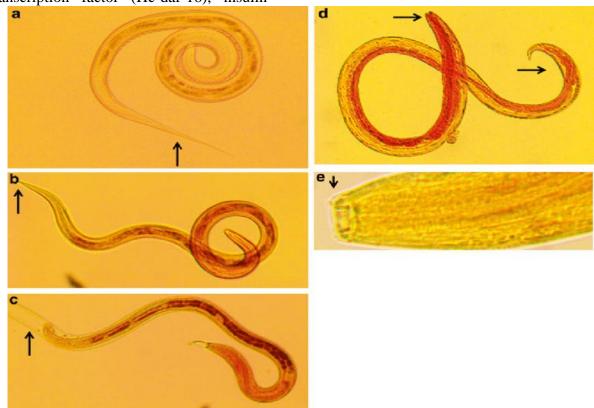
Haemonchus contortus may evolve diapause while on an infective-free-living L3s (iL3s), or beginning of the L4 (Fig. 3) of the nematode stage in order to safeguard it against a harsh environment (32). Knowing this transformation could reveal major molecules as new drug attractions provide and new insights into protection and management (33). The L3 (iL3s)-based nematodes have been proven have to higher common phenotypic, behavioral, and biological characteristics with the longof standing phase free-living Caenorhabditis elegans (34).The "dauer hypothesis" considers that the iL3 is comparable to that of the dauer stage of C. elegans in terms of developmental and

www.qu.edu.iq/journalvm



functional applications and controlled by a relatively similar method (35). The insulinlike component regulates the long-term development of C. elegans preserved in many parasitic species (35). The insulinlike signaling pathway genes are fork head transcription factor (Hc-daf-16), insulin

receptor (Hc-daf-2), phosphoinositide 3-kinases (PI3Ks) component catalytic subunit (Hc-age-1), regulatory subunit (Hc-aap-1), and phosphoinositide dependent protein kinase-1 gene (Hc-pdk-1) (32–35).



Fifure(3): The Larva shape of Haemonchus contortus https://www.researchgate.net/publication/266152011 Evaluation of the Role of Galectins in Parasite Immunity/figures?lo=1

They are intestinal genes which regulate the growth of the nematodes (36). It was found that C. elegans daf-2 mutant can be rescued by Hc-daf-2, which means that similar functions of Hc-daf-2 to Cedaf-2. PI3K kinase, Hc-age-1, and Hc-aap-1 are the downstream of Hc-DAF-2. The C. elegans age-1-mutant can not be rescued by the Hc-age-1 like Ce-age-1 (36). RNAseq analysis shows that these four genes have the maximum amount of transcription expression in iL3, which displayed that these genes could play a significant role in controlling iL3 against an unpleasant surrounding environment (36). In summary, the results from these experiments indicate that insulin-like

signals are functionally conserved between the two worms. In contrast, a bioinformatic analysis further supports the mapped insulin-like signal route of transcriptomic and genetic information for this parasite (36).

Besides, researchers are actively working on other signals such as a TGF beta signal pathway that is important for the normal growth of C. elegans, and their function in H. contortus diapause in iL3 must be understood (37). Besides being in a position to protect themselves from drying, H. contortus iL3s have also established genes that participate in this biological cycle (38). Two versions of the major gene transcript were called Hc-ubq

www.qu.edu.iq/journalvm



and Hc-gst based on their homologous ubiquitin in C. elegans and glutathione-Stransferase in H. contortus, respectively (37,38). The silencing in L3 of H. contortus of Hc-ubq or Hc-gst by RNAi decreased the rate of survival, leading to its resistance (37). The Hc38 silencing, first observed with the blot of Northern and expressed in the intestinal strongly microvilli by in situ position, decreased by 50 and 48.6 percent, respectively by taking iL3 from H. contortus into dsRNA, which utilized to invade sheep (37,38).

The L4 has short body, low metabolic rate, and crystal rod-like in the intestinal canal to safeguard them in the cold niches (39). Gene expression includes Hc-daf-22 (3-ketoacyl-CoA thiolase), Hc-maoc-1 (enoyl-CoA hydratase), and Hc-hsp-70 (heat-shock protein 70). Hc-daf-22 and Hc-maoc-1 have similarities in C. Ce-daf-22 and Ce-maoc-1, elegans, respectively, and may act in the growth of the nematode and its peroxisomal βoxidation (40).The Hc-hsp-70 overexpression manages the reduction of hsp-1 in C. elegans, indicating similar activity to that of the Ce-hsp-1 that helps of parasitic survival and invasion. Hc-fau, human fau and C. elegans rps30 analogs (ribosomal protein S30 coder), with a conserved domain of the S30 (expressed in the nucleus) and a diverged ubiquitin-like (UBiL) protein domain (cytoplasm-located expression) (41). They act in the egg-laying process and the life-span range of the C. elegans, indicating potential activities diapause based regulation in H. contortus (42).

Vaccination

Increasing consumption of anthelmintics for the care and mitigation of human and animal parasitic nematode diseases has created significant problems with anthelmintic and pharmaceutical resistance around the world (43). The vaccine is an effective method to manage parasitic nematodes, like H. contortus (44). Remarkable improvement identification of several possible antigens from H. contortus over the past two decades has been done in order to enhance influential immune thresholds in the (43,44). Here, immune system recombinant subunit vaccine and DNA vaccination have been synthesized the properties and defensive molecular potency of the key-antigens (44).

Recombinant gene subunit-based vaccines

In terms of the production of consumer vaccines, special attention has been given gene recombination, an essential to biological innovation (14).**Partial** safeguards with H. contortus recombinant antigens has been granted for immunization (45). A 110kDa central membrane glycoprotein complex was the strongest described nominee for the H. contortus antigen named H11 (14,45). The immunogen (> 90% decrease in fecal egg counts (FECs), > 75% lowering of the weight on a worm) (45). Consequently, some scientists anticipate the recombinant vaccine for potential large-scale development to be produced to promote the realistic usage of the hosts for immune defense (14). Escherichia coli had H11-1 and H11-2 variant expression, which had slightly higher aminopeptidase activity than H11-1 with enzyme activity (14,45). It has been inoculated twice with H11-1. H11-2 and H11-1 and H11-2 mixed with phosphate buffered saline. **Partial** protection has been afforded through immunization with H11-1 and H11-2 blends (29% reduction in FECs and 18% lowering of worm load) (14,45). The three (H11-1, -2, and -3) fragments of H11 gene have also been inserted in the vector for yeast expression and the lithium chloride

www.qu.edu.iq/journalvm



process used to convert recombinant Pichia Pastoris X-33. RT-PCR identified transcriptions and SDS-PAGE, and Western blot verified glycosylation of the proteins. Even so, the recombinant molecule containing three isoforms of H11 did not show protection (14,45).

As a medium, Celegans were also attempted to convey H11 to boost safety apart from E. coli and P. pastoris. (46) In this experiment, a flanking area of 1517bp 5" and part of the first H11 exon. Celegans are subcloned into the upstream region of the pPD95,77 vector, the contortus and homologous genes of the C. elegans were respectively sub-clones (47). Microinjected into the C. elegans gonads, respectively. The findings revealed numerous trends of transcriptional expression powered by freeliving and blood-sucking nematodes from their promoters, demonstrating heterogeneous supply of C. elegans (46). The trans-HPS recombinant has been developed (a 1710 bp isoform gene fragment of the H11 gene) (46).Vaccination from transgenic worms with crude Trans-HPS led to a decrease in the FEC's by 38% and a decrease in worm weight by 25% (46,47). However, E. coli presented that in the immunization studies sheep were not secured by a gene fragment from 670 bp to 1710 bp isoform H11. In addition to possible antigen H11, the recombinant galectin Hco-gal-m / f (obtained respectively from male and female nematodes) is expressed from H. contortus in E. coli Vaccinations with 200 ug protein reduced 48-46 percent fecal egg output, with Freund's adjuvant. Vaccination with a mixture of Hco-gal-m / f recombinant proteins has a function to play in the defense of goats (46,47).

DNA-based vaccines

DNA vaccination is a bioengineered DNA defense strategy that allows cells to actively produce antigens and vaccinated animals to develop safe immune responses

to diseases (48). In contrast with traditional vaccines, DNA vaccines have several benefits, including a broader variety of specific immune responses (48). This is a new approach to managing infectious For the illustration. parasites. injection of antigen-specific immune-based plasmid codes for exogenous antigenic release (48). Several observations of partial safety in goats after DNA vaccination were reported and identified. Immunization of goats between 8 and 10 months of age, with HC29, DNA encoder (48). Different antibodies and partially immune defense declines in FECs and (36% worm comparison burdens), to goats who obtained just PBS, have been caused by the GPX (contortus glutathione peroxidase) (48). Vaccination with DNA Vaccines, comprising three fragments, encoding sections for H11-1 and caprine interleukin 2 (IL-2), led to a significant reduction of the fecal output, the abomasal worm and lymphocytes by 57 and 47 percent of specific serum immunoglobulins G (IgG), IgA, CD4 + T lymphocytes and CD8 + T lymphocytes (48).

In H. contortus, Glyceraldehyde-3phosphate dehydrogenase, as a protection against experimental infections of H. contortus in goats was evaluated with vaccine The DNA (49).analysis indicates that the vaccinated adds essential qualification peripheral and local mucosal immune responses and supported the development of lymphocytes of CD4+T and B but supplied only limited protection in comparison with control groups (35% decrease in FECs, and 38% decrease in the worm weight) (49). DNA vaccine may be much more secure (46% decrease in FECs and 51% decrease in worm burden) against the resulting disease in the goats through vaccination of disorganized muscle family member (Dim-1) (49).

The processes recognition of the immune control in order to identify useful vaccine antigens. Several experiments

www.qu.edu.iq/journalvm



performed during parasite were a invasion to examine the function of immune suppression by studying the interplay of parasite and host cell molecules (50).To achieve so, venipuncture has isolated the target peripheral blood mononuclear cells (PBMCs) of goat blood. PBMC is the mixture of activated cell subpopulations, primarily lymphocytes, monocytes and dendritic cells (T cells, B cells and NK cells) (50). The process for communicating host cells with several essential parasite molecules is crucial for subpopulation. For instance. The Hco-galm / f (rHco-gal-m / f) recombinant Contortus Galectin peptides are grown using PBMC of caprines and investigated in the PBMC the impact of rHco-gal-m / fon the induction of apoptosis (50). The pathways underlying immunomodulation caused by rHco-gal-m / f were also studied together in a proteomic and transcriptomic way (50). PBMC. The results indicate that rHco-gal-m/f can bind to the surface of the PBMC and serve as an inflammatory reaction to ease H. contortus immune escape (50).

Based on the analysis, two H-co and F binding partners, Transmembrane Protein (TMEM147), and Transmembrane Protein 63A (TMEM63A) were also discovered through further yeast twohybrid testing (51). The communication between galectin and TMEM147 mainly mediates cell division, death of cells, and the transcription of cytokine in the PBMC Along with TMEM63A, (51).membranous protein participates in the control of galectin and the development of the PBMC in the phagocytosis and nitric oxides. Even so, in the migratory galectin control and IFN-B transcript of goat PBMC, TMEM63A performs a significant role than TMEM147 (51). These research results provide a unique view for the clarification by nematodes and parasitehost interaction of the processes involved in immune avoidance (51).

As a vaccine nominee, H. contortus excretory and secretory items include different proteins that may activate or suppress the host's immune system and are implicated in the pathophysiology of worms (52). Work suggested that the PBMC in vitro is suppressed by H. contortus excretory and secretory items. The items were used to inhibit IL-4, IFN-α supply, boost cytokine suppressive IL-10, strengthen inflammatory modulator IL-17, and repress the manufacturing of the nitric oxide (52). Protein engagement with the PBMC in vivo using liquid chromatography/tandem mass spectrometry from these materials in various developmental stages showed a particles were detected of 407 interacting with the PBMC, 14-3-3 protein in all parasites as PBMC-interacting proteins (52). IL-4 output declined, and the PBMC in vitro proliferation repressed by the 14-3-3 isoform 2 (rHcftt-2) (52).

In addition, an important immunogenic component was also observed in 24 kDa H. contortus excretory / secretary proteins (53). The immune interactions between HcES-24 and Goat PBMC recombinant protein have shown an improvement in IL 4, IL-10, IL-17, and cell migration. Nonetheless, PBMC multiplication and NO development were greatly inhibited by the relationship (53). The results showed that the rHcES-24 had significant regulatory impacts on the PBMC (53).

Conclusion

The use of molecular-based approaches is critical in identifying the resistance type and its mechanism for the best utilization of therapeutic drugs against *Haemonchus contortus*.

References

1. Jones JT, Haegeman A, Danchin EGJ, Gaur HS, Helder J, Jones MGK, et al. Top 10 plant-

www.qu.edu.iq/journalvm



- parasitic nematodes in molecular plant pathology. Mol Plant Pathol [Internet]. 2013 Dec [cited 2020 Aug 16];14(9):946–61. Available from: https://pubmed.ncbi.nlm.nih.gov/23809086/
- O'Connor LJ, Walkden-Brown SW, Kahn LP. Ecology of the free-living stages of major trichostrongylid parasites of sheep. Vet Parasitol [Internet]. 2006 Nov 30 [cited 2020 Aug 16];142(1-2):1-15. Available from: https://pubmed.ncbi.nlm.nih.gov/17011129/
- 3. Blaxter M, Koutsovoulos G. The evolution of parasitism in Nematoda. Parasitology [Internet]. 2015 Feb 10 [cited 2020 Aug 16];142(Suppl 1):S26–39. Available from: https://pubmed.ncbi.nlm.nih.gov/24963797/
- 4. Bundy DAP, Appleby LJ, Bradley M, Croke K, Hollingsworth TD, Pullan R, et al. 100 Years of Mass Deworming Programmes: A Policy Perspective From the World Bank's Disease Control Priorities Analyses. In: Advances in Parasitology [Internet]. Academic Press; 2018 [cited 2020 Aug 16]. p. 127–54. Available from: https://pubmed.ncbi.nlm.nih.gov/29753337/
- 5. McKellar QA, Jackson F. Veterinary anthelmintics: Old and new. Trends Parasitol [Internet]. 2004 Oct [cited 2020 Aug 16];20(10):456–61. Available from: https://pubmed.ncbi.nlm.nih.gov/15363438/
- Kaplan RM, Vidyashankar AN. An inconvenient truth: Global worming and anthelmintic resistance. Vet Parasitol [Internet]. 2012 [cited 2020 Aug 16];186(1–2):70–8. Available from: https://pubmed.ncbi.nlm.nih.gov/22154968/
- 7. Hay SI, Abajobir AA, Abate KH, Abbafati C, Abbas KM, Abd-Allah F, et al. Global, regional, and national disability-adjusted life-years (DALYs) for 333 diseases and injuries and healthy life expectancy (HALE) for 195 countries and territories, 1990-2016: A systematic analysis for the Global Burden of Disease Study 2016. Lancet [Internet]. 2017 Sep 16 [cited 2020 Aug 16];390(10100):1260–344. Available from: https://pubmed.ncbi.nlm.nih.gov/28919118/
- 8. Hewitson JP, Maizels RM. Vaccination against helminth parasite infections. Expert Rev Vaccines [Internet]. 2014 [cited 2020 Aug 16];13(4):473–87. Available from: https://pubmed.ncbi.nlm.nih.gov/24606541/
- 9. Sallé G, Laing R, Cotton JA, Maitland K, Martinelli A, Holroyd N, et al. Transcriptomic

- profiling of nematode parasites surviving vaccine exposure. Int J Parasitol [Internet]. 2018 Apr 1 [cited 2020 Aug 16];48(5):395–402. Available from: https://pubmed.ncbi.nlm.nih.gov/29534987/
- 10. Laing R, Kikuchi T, Martinelli A, Tsai IJ, Beech RN, Redman E, et al. The genome and transcriptome of Haemonchus contortus, a key model parasite for drug and vaccine discovery. Genome Biol [Internet]. 2013 Aug 28 [cited 2020 Aug 16];14(8). Available from: https://pubmed.ncbi.nlm.nih.gov/23985316/
- 11. Doyle SR, Laing R, Bartley DJ, Britton C, Chaudhry U, Gilleard JS, et al. A Genome Resequencing-Based Genetic Map Reveals the Recombination Landscape of an Outbred Parasitic Nematode in the Presence of Polyploidy and Polyandry. Genome Biol Evol [Internet]. 2018 Feb 1 [cited 2020 Aug 16];10(2):396–409. Available from: https://pubmed.ncbi.nlm.nih.gov/29267942/
- 12. Gilleard JS. Haemonchus contortus as a paradigm and model to study anthelmintic drug resistance. Parasitology [Internet]. 2013 Oct [cited 2020 Aug 16];140(12):1506–22. Available from: https://pubmed.ncbi.nlm.nih.gov/23998513/
- 13. Coghlan A, Tyagi R, Cotton JA, Holroyd N, Rosa BA, Tsai IJ, et al. Comparative genomics of the major parasitic worms. Nat Genet [Internet]. 2019 Jan 1 [cited 2020 Aug 16];51(1):163–74. Available from: https://pubmed.ncbi.nlm.nih.gov/30397333/
- 14. Wang C, Li F, Zhang Z, Yang X, Ahmad AA, Li X, et al. Recent research progress in China on Haemonchus contortus. Front Microbiol [Internet]. 2017 Aug 24 [cited 2020 Aug 16];8(AUG):1509. Available from: /pmc/articles/PMC5574212/?report=abstract
- 15. Kwa MSG, Veenstra JG, Roos MH. Benzimidazole resistance in Haemonchus contortus is correlated with a conserved mutation at amino acid 200 in β-tubulin isotype 1. Mol Biochem Parasitol [Internet]. 1994 [cited 2020 Aug 16];63(2):299–303. Available from: https://pubmed.ncbi.nlm.nih.gov/7911975/
- 16. Prichard RK. Genetic variability following selection of Haemonchus contortus with anthelmintics. Trends Parasitol [Internet]. 2001 Sep 1 [cited 2020 Aug 16];17(9):445–53. Available from: https://pubmed.ncbi.nlm.nih.gov/11530357/
- 17. Rufener L, Kaminsky R, Mäser P. In vitro

www.qu.edu.iq/journalvm



- selection of Haemonchus contortus for benzimidazole resistance reveals a mutation at amino acid 198 of β -tubulin. Mol Biochem Parasitol [Internet]. 2009 [cited 2020 Aug 16];168(1):120–2. Available from: https://pubmed.ncbi.nlm.nih.gov/19616042/
- 18. Kwa MSG, Veenstra JG, Van Dijk M, Roos MH. β-Tubulin genes from the parasitic nematode Haemonchus contortus modulate drug resistance in Caenorhabditis elegans. J Mol Biol [Internet]. 1995 Mar 3 [cited 2020 Aug 16];246(4):500–10. Available from: https://pubmed.ncbi.nlm.nih.gov/7877171/
- 19. Silvestre A, Cabaret J. Mutation in position 167 of isotype 1 β-tubulin gene of Trichostrongylid nematodes: Role in benzimidazole resistance? Mol Biochem Parasitol [Internet]. 2002 Apr 9 [cited 2020 Aug 16];120(2):297–300. Available from: https://pubmed.ncbi.nlm.nih.gov/11897135/
- 20. Ghisi M, Kaminsky R, Mäser P. Phenotyping and genotyping of Haemonchus contortus isolates reveals a new putative candidate mutation for benzimidazole resistance in nematodes. Vet Parasitol [Internet]. 2007 Mar 31 [cited 2020 Aug 16];144(3–4):313–20. Available from: https://pubmed.ncbi.nlm.nih.gov/17101226/
- 21. Zhang Z, Gasser RB, Yang X, Yin F, Zhao G, Bao M, et al. Two benzimidazole resistance-associated SNPs in the isotype-1 β-tubulin gene predominate in Haemonchus contortus populations from eight regions in China. Int J Parasitol Drugs Drug Resist [Internet]. 2016 Dec 1 [cited 2020 Aug 16];6(3):199–206. Available from: https://pubmed.ncbi.nlm.nih.gov/27760394/
- 22. Roeber F, Jex AR, Gasser RB. Advances in the diagnosis of key gastrointestinal nematode infections of livestock, with an emphasis on small ruminants. Biotechnol Adv [Internet]. 2013 Dec [cited 2020 Aug 16];31(8):1135–52. Available from: https://pubmed.ncbi.nlm.nih.gov/23376340/
- 23. Kotze AC, Prichard RK. Anthelmintic Resistance in Haemonchus contortus. History, Mechanisms and Diagnosis. Adv Parasitol [Internet]. 2016 [cited 2020 Aug 16];93:397–428. Available from: https://pubmed.ncbi.nlm.nih.gov/27238009/
- 24. Redman E, Sargison N, Whitelaw F, Jackson F, Morrison A, Bartley DJ, et al. Introgression of ivermectin resistance genes into a susceptible Haemonchus contortus strain by

- multiple backcrossing. PLoS Pathog [Internet]. 2012 Feb [cited 2020 Aug 16];8(2). Available from:
- https://pubmed.ncbi.nlm.nih.gov/22359506/
- 25. Luo X, Shi X, Yuan C, Ai M, Ge C, Hu M, et al. Genome-wide SNP analysis using 2b-RAD sequencing identifies the candidate genes putatively associated with resistance to ivermectin in Haemonchus contortus. Parasites and Vectors [Internet]. 2017 Jan 17 [cited 2020 Aug 16];10(1). Available from: https://pubmed.ncbi.nlm.nih.gov/28095895/
- 26. Lespine A, Ménez C, Bourguinat C, Prichard RK. P-glycoproteins and other multidrug resistance transporters in the pharmacology of anthelmintics: Prospects for reversing transport-dependent anthelmintic resistance. Int J Parasitol Drugs Drug Resist [Internet]. 2012 Dec [cited 2020 Aug 16];2:58–75. Available from: https://pubmed.ncbi.nlm.nih.gov/24533264/
- 27. Khan S, Nisar A, Yuan J, Luo X, Dou X, Liu F, et al. A whole genome re-sequencing based GWA analysis reveals candidate genes associated with ivermectin resistance in Haemonchus contortus. Genes (Basel) [Internet]. 2020 Apr 1 [cited 2020 Aug 16];11(4). Available from: /pmc/articles/PMC7230667/?report=abstract
- 28. Laing R, Maitland K, Lecová L, Skuce PJ, Tait A, Devaney E. Analysis of putative resistance gene loci in UK field populations of Haemonchus contortus after 6 years of macrocyclic lactone use. Int J Parasitol [Internet]. 2016 Sep 1 [cited 2020 Aug 16];46(10):621–30. Available from: /pmc/articles/PMC5011429/?report=abstract
- 29. Alam MBB, Omar AI, Faruque MO, Notter DR, Periasamy K, Mondal MMH, et al. Single nucleotide polymorphisms in candidate genes are significantly associated with resistance to Haemonchus contortus infection in goats. J Anim Sci Biotechnol [Internet]. 2019 Dec 15 [cited 2020 Aug 16];10(1):30. Available from:
 - https://jasbsci.biomedcentral.com/articles/10.11 86/s40104-019-0327-8
- 30. Becker GM, Davenport KM, Burke JM, Lewis RM, Miller JE, Morgan JLM, et al. Genome-wide association study to identify genetic loci associated with gastrointestinal nematode resistance in Katahdin sheep. Anim Genet [Internet]. 2020 Mar 1 [cited 2020 Aug 16];51(2):330–5. Available from:

www.qu.edu.iq/journalvm



/pmc/articles/PMC7064973/?report=abstract

- Luo X, Shi X, Yuan C, Ai M, Ge C, Hu M, et al. Genome-wide SNP analysis using 2b-RAD sequencing identifies the candidate genes putatively associated with resistance to ivermectin in Haemonchus contortus. Parasit Vectors [Internet]. 2017 Dec 17 [cited 2020 Aug 16];10(1):31. Available http://parasitesandvectors.biomedcentral.com/art icles/10.1186/s13071-016-1959-6
- 32. Li FC, Gasser RB, Lok JB, Korhonen PK, He L, Di W Da, et al. Molecular characterization of the Haemonchus contortus phosphoinositide-dependent protein kinase-1 gene (Hc-pdk-1). Parasites and Vectors [Internet]. 2016 Feb 3 [cited 2020 Aug 17];9(1). Available from: https://pubmed.ncbi.nlm.nih.gov/26842781/
- Li FC, Gasser RB, Lok JB, Korhonen PK, 33. Wang YF, Yin FY, et al. Exploring the role of two interacting phosphoinositide 3-kinases of Haemonchus contortus. Parasites and Vectors [Internet]. 2014 [cited 2020 Aug 17];7(1). from: https://pubmed.ncbi.nlm.nih.gov/25388625/
- Li F, Lok JB, Gasser RB, Korhonen PK, Sandeman MR, Shi D, et al. Hc-daf-2 encodes an insulin-like receptor kinase in the barber's pole worm, Haemonchus contortus, and restores partial dauer regulation. Int J Parasitol [Internet]. 2014 [cited 2020 Aug 17];44(7):485-Available 96 from: https://pubmed.ncbi.nlm.nih.gov/24727120/
- Hu M, Lok JB, Ranjit N, Massey HC, Sternberg PW, Gasser RB. Structural and functional characterisation of the fork head transcription factor-encoding gene, Hc-daf-16, from the parasitic nematode Haemonchus contortus (Strongylida). Int J Parasitol [Internet]. 2010 Mar 15 [cited 2020 Aug 17];40(4):405–15. Available from: https://pubmed.ncbi.nlm.nih.gov/19796644/
- Mohandas N, Hu M, Stroehlein AJ, Young ND, Sternberg PW, Lok JB, et al. Reconstruction of the insulin-like signalling pathway of Haemonchus contortus. Parasites and Vectors [Internet]. 2016 Feb 3 [cited 2020 17];9(1):64. Available from: https://pubmed.ncbi.nlm.nih.gov/26842675/
- Hartman D, Donald DR, Nikolaou S, Savin KW, Hasse D, Presidente PJA, et al. Analysis of developmentally regulated genes of the parasite Haemonchus contortus. Int J Parasitol [Internet]. 2001 [cited 2020 Aug

- 17];31(11):1236–45. Available from: https://pubmed.ncbi.nlm.nih.gov/11513893/
- Yang Y, Ma Y, Chen X, Guo X, Yan B, 38. Du A. Screening and analysis of Hc-ubq and Hc-gst related to desiccation survival of infective Haemonchus contortus larvae. Vet Parasitol [Internet]. 2015 Jun 15 [cited 2020 Aug 17];210(3–4):179–85. Available from: https://pubmed.ncbi.nlm.nih.gov/25913452/
- Guo X, Zhang H, Zheng X, Zhou Q, Yang 39. Y, Chen X, et al. Structural and functional characterization of a novel gene, Hc-daf-22, from the strongylid nematode Haemonchus contortus. Parasites and Vectors [Internet]. 2016 Jul 29 [cited 2020 Aug 17];9(1). Available from:
 - https://pubmed.ncbi.nlm.nih.gov/27472920/
- Ding H, Shi H, Shi Y, Guo X, Zheng X, Chen X, et al. Characterization and function analysis of a novel gene, Hc-maoc-1, in the parasitic nematode Haemonochus contortus. Parasites and Vectors [Internet]. 2017 Feb 6 [cited 2020 Aug 17];10(1):1–14. Available
 - https://pubmed.ncbi.nlm.nih.gov/28166831/
- 41. Zhang H, Zhou Q, Yang Y, Chen X, Yan B, Du A. Characterization of heat shock protein 70 gene from Haemonchus contortus and its expression and promoter analysis Caenorhabditis elegans. Parasitology [Internet]. 2013 May [cited 2020 Aug 17];140(6):683-94. Available from: https://pubmed.ncbi.nlm.nih.gov/23360558/
- Yan B, Guo X, Zhou Q, Yang Y, Chen X, Sun W, et al. Hc-fau, a novel gene regulating diapause in the nematode parasite haemonchus contortus. Int J Parasitol [Internet]. 2014 Oct 1 [cited 2020 Aug 17];44(11):775-86. Available
 - https://pubmed.ncbi.nlm.nih.gov/25058511/
- Tak IR, Dar JS, Dar SA, Ganai BA, Chishti MZ, Ahmad F. A comparative analysis of various antigenic proteins found in Haemonchus contortus--a review. Mol Biol (Mosk) [Internet]. 2015 Nov 1 [cited 2020 Aug 17];49(6):883–90. Available from: https://pubmed.ncbi.nlm.nih.gov/26710767/
- Bassetto CC, Amarante AFT. Vaccination of sheep and cattle against haemonchosis. J Helminthol [Internet]. 2015 Sep 7 [cited 2020 17];89(5):517–25. Available from: https://pubmed.ncbi.nlm.nih.gov/25891536/
- Newton SE, Munn EA. The development 45.

www.qu.edu.iq/journalvm



- of vaccines against gastrointestinal nematode parasites, particularly Haemonchus contortus. Parasitol Today [Internet]. 1999 Mar 1 [cited 2020 Aug 17];15(3):116–22. Available from: https://pubmed.ncbi.nlm.nih.gov/10322325/
- 46. Yanming S, Ruofeng Y, Muleke CI, Guangwei Z, Lixin X, Xiangrui L. Vaccination of goats with recombinant galectin antigen induces partial protection against Haemonchus contortus infection. Parasite Immunol [Internet]. 2007 Jun [cited 2020 Aug 17];29(6):319–26. Available from: https://pubmed.ncbi.nlm.nih.gov/17518950/
- 47. Zhou QJ, Yang Y, Guo XL, Duan LJ, Chen XQ, Yan BL, et al. Expression of Caenorhabditis elegans-expressed Trans-HPS, partial aminopeptidase H11 from Haemonchus contortus. Exp Parasitol [Internet]. 2014 [cited 2020 Aug 17];145(1):87–98. Available from: https://pubmed.ncbi.nlm.nih.gov/25128369/
- 48. Zhao GW, Yan RF, Muleke CI, Sun YM, Xu LX, Li XR. Vaccination of goats with DNA vaccines encoding H11 and IL-2 induces partial protection against Haemonchus contortus infection. Vet J [Internet]. 2012 Jan [cited 2020 Aug 17];191(1):94–100. Available from: https://pubmed.ncbi.nlm.nih.gov/21330170/
- 49. Yan R, Wang J, Xu L, Song X, Li X. DNA vaccine encoding Haemonchus contortus Actin induces partial protection in Goats. Acta Parasitol [Internet]. 2014 Oct 1 [cited 2020 Aug 17];59(4):698–709. Available from: https://pubmed.ncbi.nlm.nih.gov/25236283/

- 50. Wang W, Yuan C, Wang S, Song XK, Xu LX, Yan RF, et al. Transcriptional and proteomic analysis reveal recombinant galectins of Haemonchus contortus down-regulated functions of goat PBMC and modulation of several signaling cascades in vitro. J Proteomics [Internet]. 2014 Feb 26 [cited 2020 Aug 17];98:123–37. Available from: https://pubmed.ncbi.nlm.nih.gov/24401599/
- 51. Li Y, Yuan C, Wang L, Lu M, Wang Y, Wen Y, et al. Transmembrane protein 147 (TMEM147): Another partner protein of Haemonchus contortus galectin on the goat peripheral blood mononuclear cells (PBMC). Parasites and Vectors [Internet]. 2016 Jun 23 [cited 2020 Aug 17];9(1). Available from: https://pubmed.ncbi.nlm.nih.gov/27337943/
- 52. Gadahi JA, Ehsan M, Wang S, Zhang ZC, Wang Y, Yan RF, et al. Recombinant protein of Haemonchus contortus 14-3-3 isoform 2 (rHcftt-2) decreased the production of IL-4 and suppressed the proliferation of goat PBMCs in vitro. Exp Parasitol [Internet]. 2016 Dec 1 [cited 2020 Aug 17];171:57–66. Available from: https://pubmed.ncbi.nlm.nih.gov/27751769/
- 53. Gadahi JA, Li B, Ehsan M, Wang S, Zhang Z, Wang Y, et al. Recombinant Haemonchus contortus 24 kDa excretory/ secretory protein (rHcES-24) modulate the immune functions of goat PBMCs in vitro. Oncotarget [Internet]. 2016 [cited 2020 Aug 17];7(51):83926–37. Available from: https://pubmed.ncbi.nlm.nih.gov/27893414/