STUDY THE PROTECTIVE ROLE OF QUERCETIN AGAINST O-ANISIDINE TOXICITY ON SOME HEMATOLOGICAL PARAMETERS OF LABORATORY MALE RATS (*RATTUS NORVEGICUS*).

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ABSTRACT

The study was designed to assess the protective role of Quercetin against Oanisidine toxicity. 24 male rats (Rattus norvegicus) were used and divided into 3 equal groups of 8 male rats each. The first group was the control group in which the animals were fed on a standard ration for 15 days, then they were terminated. The second group was fed on a ration contains 1000mg/kg O-anisidine hydrochloride for 15.The third group was fed on a ration contains 1000mg/kg O-anisidine hydrochloride + 80mg/kg Quercetin dihydrate for 15. The results revealed that the treatment with Oanisidine hydrochloride for 15 days (1rst treated group) caused significant decrease in the R.B.C. count, Hb concentration, P.C.V. percentage, neutrophil and lymphocyte counts and it caused significant increase in platelets count, total leukocytes, monocytes, eosinophil and basophil counts, as compared with control group. When Quercetin dihydrate was offered as a protective agent in the ration of the 2nd treated group, it showed a significant ameliorating effect by increasing the R.B.C. count, Hb concentration, P.C.V. percentage, neutrophil and lymphocyte counts and it caused significant decrease in platelets count, total leukocytes, monocytes, eosinophil and basophil counts, as compared with the 1rst treated group. For the blood indices (RDW, MCH, MCHC, MCV, and MPV) there were no significant differences among all the experiment groups except for the mean platelet volume (MPV), where Oanisidine hydrochloride caused significant decrease in the MPV of the 1rst treated group as compared with control and 2nd treated group at ($P \le 0.05$).

INTRODUCTION

Ortho-anisidine is one of the widely spread family known as aromatic amines (1). (2) mentioned that the principal commercial use of O-anisidine is an intermediate in the manufacture of dyes and pharmaceuticals and as an intermediate in the manufacture of synthetic guaiacol and its derivatives. It has been known that Oanisidine is released from textiles and leather goods colored with azo dyes and a large part of the population may be exposed and as a constituent of cigarette smoke (3). Hemoglobin adducts of O-anisidine were detected in blood samples of persons living in urban or rural areas of Germany (4). (5) mentioned that young children are exposed by oral suckling clothes which are colored with O-anisidine based dyes, and workplace exposure occurs by inhalation in the production and processing of Oanisidine, dermal contact during the formulation of O-anisidine based printing inks, general population in dermal contact with packing materials printed with O-anisidine based pigments; and man exposed indirectly via the environment. Flavonoids are a group of naturally occurring polyphenolic compounds widely distributed as secondary metabolites in plant kingdom (6). Quercetin (3, 5, 7, 3, 4-pentahydroxy flavon), is one of the most prominent dietary antioxidants (7). Quercetin occurs in glycosylated form in French beans, broccoli, and apples and especially in onions (8). Quercetin is the most abundant antioxidant in the nature and has an antioxidant potential four times that of vitamin E (9). Quercetin has been reported to increase the genomic stability in rats and enhance the antioxidative defense system by up regulating antioxidant enzymes (10). It has many beneficial effects in human health, including cardiovascular protection, anticancer activity, anti-ulcer effects, anti-allergy activity, cataract prevention, antiviral activity and anti-inflammatory effects (11). Quercetin prevents oxidant injury and cell death by several mechanisms, such as scavenging oxygen radicals, protecting against lipid peroxidation and chelating metal ions (12).

MATERIALS AND METHODS

Experimental animals and diets

The experiment was conducted at the animal house of the Veterinary Medicine College–University of Basra, where 24 male rats (*Rattus norvegicus*) of 170–175 grams weights were used. The experiment conditions were unified for all animals, where the room temperature was set between 20–25 C by the use of an air

conditioner, and the daily light period was 12 hours by the use of two fluorescent lamps, and the humidity rate was about 50 %. Food and water were provided daily (*ad libitum*).

Experimental design

The animals were allocated into three groups; each group consists of 8 male rats. The hematological tests were done after 15 days of the experiment. The groups were:

Control group: In this group, 8 male rats were fed with standard ration for 15 days then they were terminated for the necessary tests.

The first treated group (T1) group: This group consisted of 8 male rats which were fed with a ration contains O-Anisidine hydrochloride 1000mg/kg for 15 days, then they were terminated for the necessary tests.

The second treated group (T2) group: This group consisted of 8 male rats which were fed with a ration contains 1000mg/kg O-Anisidine hydrochloride +80 mg/kg Quercetin dihydrate for 15 days, then they were terminated for the necessary tests.

Specimens' collection

After the animals were anaesthetized by holding them apart in a chloroform containing conduit. After their chests were dissected, blood samples were collected via cardiac puncture by using 5ml disposable syringe according to the method of (13). Then the blood was put in an anticoagulant containing tubes.

Hematological parameters

All the hematological parameters in this study were obtained by the use of a highly developed hematology analyzer (Ruby[®], Germany made), where 1ml of the noncoagulated blood is aspirated by a special needle equipped within the device and then the analyzed results would appear in a detailed printed report.

Statistical Analysis

In this study, ANOVA Analysis and LSD tests are used according to (SPSS version 18) programme at the (P \leq 0.05) to find the significant differences among means for all treatments.

RESULTS AND DISCUSSION

The results revealed that the treatment with O-anisidine hydrochloride for 15 days (T1 group) caused significant decrease in the R.B.C. count, Hb concentration, P.C.V. percentage, neutrophil and lymphocyte counts and it caused significant increase in platelets count, total leukocytes, monocytes, eosinophil and basophil counts, as compared with control group at (P \leq 0.05) tables (1,3). When Quercetin dihydrate was offered as a protective agent in the ration of the (T2 group), it showed a significant ameliorating effect by increasing the R.B.C. count, Hb concentration, P.C.V. percentage, neutrophil and lymphocyte counts and it caused significant decrease in platelets count, total leukocytes, monocytes, eosinophil and basophil counts, as compared with the T1 group at (P \leq 0.05) tables (1, 3). For the blood indices (RDW, MCH, MCHC, MCV, and MPV) there were no significant differences among all the experiment groups except for the mean platelet volume (MPV), where O-anisidine hydrochloride caused significant decrease in the MPV of the T1 group as compared with control and T2 group at (P \leq 0.05) table (2).

These deleterious effects of O-anisidine on the blood parameters can be explained pending upon either the direct effect of O-anisidine or indirectly by the effects of the metabolites which are resulted from its metabolism in the liver of rat. Where, Oanisidine is oxidized by human, rat and rabbit hepatic microsomes containing cytochromes P450 not only to N-(2-methoxyphenyl)hydroxylamine, but that this compound is a subject of complex redox cycling reactions, forming also oaminophenol, 2- nitrosoanisole and one additional metabolite, the exact structure of which has not been identified as yet (14). O-Demethylation of 2-nitroanisole to 2nitrophenol and its hydroxylated products, 2, 5-dihydroxynitrobenzene and 2, 6dihydroxynitrobenzene, (15). N-Hydroxyarylamines can be further metabolized to Nsulfonyloxyarylamines, N-acetoxyarylamines or N-hydroxyarylamine N-glucuronide. These highly reactive intermediates are responsible for the genotoxic and cytotoxic effects (8) of this class of compounds. One of the most effects of these metabolites is the formation of reactive oxygen species (ROS) especially by the metabolite Oaminophenol like superoxide radicals or hydrogen peroxide. ROS can induce oxidative damage to the cell and can form a very stable structure by extracting electrons from other sources (17). ROS are also able to generate other forms of ROS. Superoxide can be dismutated into H2O2 and oxygen. H2O2 has the ability to form

the more damaging OH, through a combination of the Fenton and Haber-Weiss reactions (18). The ROS which are not neutralized, can target biological molecules such as DNA, lipids, proteins, and carbohydrates, which can result in cell dysfunction or cell death. Red blood cell (RBC) membranes contain lipids rich in unsaturated fatty acids. RBCs are more frequently exposed to oxygen than other body tissue and, thus, are more susceptible to oxidative damage. Erythrocytes are highly susceptible to the oxidative damage due to the high cellular concentration of oxygen and hemoglobina potentially powerful promoter for the oxidative processes (19). Invasion of the RBC membrane by peroxidants may lead to cell hemolysis. Moreover, the hemoglobin in RBCs is a strong catalyst which may initiate lipid peroxidation. In addition to lipid peroxidation, oxidants affect vital –SH groups of proteins which are highly active and may be targeted during oxidative stress (20). Platelets, a major player in thrombus formation, are obviously a prime target for oxidants produced or released in the vascular lumen (21). So, the elevated platelets count in the results of our study can be explained by the following mechanisms: Production of hydrogen peroxide seems to promote thromboxane synthesis, and hence platelet aggregation, in response to arachidonic acid stimulation. This conclusion is also supported by data obtained by (22) and (23), who showed that low concentrations of hydrogen peroxide could potentiate the aggregatory response to arachidonic acid or collagen. In summary, the effects of reactive oxygen metabolites on in vitro platelet function are complex. Low levels of HO may promote thromboxane synthesis and aggregation.

Role of other platelet agonists. Another possible mechanism by which oxidants may influence platelet aggregation is through potentiation of the effects of platelet-activating factor _PAF. PAF is an autacoid released by platelets and other cell types _e.g. endothelium, leukocytes, which acts on platelets at extremely low concentrations (24). Superoxide and hydroxyl radicals can rapidly _i.e. within seconds and irreversibly inactivate plasma PAF-acetylhydrolase, the enzyme that catabolizes PAF. Once PAF is formed or released in the blood, inhibition of PAF-acetylhydrolase would enhance concentrations and prolong half-life of this powerful agonist. Thus, it may be speculated that oxygen radicals may indirectly enhance platelet aggregation, through local increases in PAF concentrations secondary to reduced breakdown of PAF. In vivo this hypothesis is indirectly supported by data derived from a study by (25), in which administration of superoxide dismutase was associated with a

significant reduction in PAF-mediated aggregation of platelets resuspended in plasma, consistent with preserved activity of plasma acetylhydrolase.

Quercetin has a high antioxidant potential, the effect that is mainly based on inactivation of reactive oxygen species (ROS). Thus, Quercetin protects the cells against free radicals (26). Quercetin, inhibit glycosylation by 52%. This is an important effect, as flavonoids are stored in RBCs (27). Pure Quercetin inhibits hemolysis, Quercetin decreases hemolysis in a dose-dependent manner. This was also shown by (28). Additionally, flavonoids have inhibitory effects on the functions of platelets and leukocytes. They also protect endothelial cells, and counterbalance the interactions between the blood stream and vascular wall, which may lead to thrombosis. The latter effect is mediated through the effect of flavonoids on human monocyte tissue factor, which itself may trigger blood coagulation (29). The mechanism of action of Quercetin has been attributed largely to the antioxidant properties, which are known to augment GSH and antioxidant enzyme levels and scavenge lipid peroxides. A concept is now emerging of "adaptogenic drugs" - drugs that increases non-specific resistance to variety of stresses. The neutropenia in the results which was caused by O-anisidine treatment came in accordance with the results obtained by (30) who found that oxidative stress caused neutropenia and accelerated neutrophil apoptosis in dogs, and also agreed with the results obtained by (31) who reported that exposure to benzene derivatives causes lymphopenia and neutropenia in rats.

Table (1). Role of Quercetin dihydrate against O-anisidine hydrochloride on
blood parameters of rats. Different letters refer to significant differences among
groups at (P≤0.05).

PARAMETERS	RBC (n×10 ⁶ /µl)	Hb (g/dl)	PCV (%)	Platelets (n×10 ³ /µl)	
GROUPS					
Control	a 8.6 ±1.1	a 14.6 ±1.2	a 44.5 ±3.4	c 542.6 ±20.6	
T1	b 6.6 ±0.9	c 10.7 ±1.2	c 33.4 ±3.8	a 876.7 ±19.62	
T2	a 7.7 ±1.0	b 12.6 ±1.7	b 38.8 ±2.3	b 586.5 ±8.6	
LSD	1.4	1.9	5.7	24.6	

Table (2). Role of Quercetin dihydrate against O-anisidine hydrochloride on blood indices of rats. Different letters refer to significant differences among groups at (P≤0.05).

PARAMETERS GROUPS	RDW (%)	MCH (pg)	MCHC (g/dl)	MPV (fl)	MCV (fl)
Control	a	a	a	a	a
	10.5	16.6	32.7	4.24	50.8
	±1.0	±1.05	±1.8	±0.22	±1.7
T1	a	a	a	b	a
	10.3	16.4	32.1	3.77	51.0
	±1.0	±1.2	±2.2	±0.8	±1.3
T2	a	a	a	a	a
	10.5	16.5	32.6	4.06	50.7
	±1.0	±1.0	±1.6	±0.24	±1.1
LSD	00	00	00	0.28	00

RARAMETERS	WBC	Nutrophil	Lympho	Acidophil	Basophil	Monocyte
GROUPS	(n×10³/µl)	(%)	(%)	(%)	(%)	(%)
Control	с	a	a	b	b	b
	6.2	12.1	80.1	1	0.3	4
	±1.2	±1.3	±1.3	±0.5	±0.07	±0.8
T1	a	b	b	a	a	a
	9.3	8.3	70.4	4	8.5	6.2
	±1.1	±1.1	±3.0	±0.6	±1.4	±0.9
T2	b	a	a	b	b	b
	7.2	11.1	77.9	0.9	1.1	4.6
	±1.3	±1.5	±4.1	±0.2	±0.8	±1.3
LSD	1.7	1.9	6.1	2.06	0.9	0.9

Table (3). Role of Quercetin dihydrate against O-anisidine hydrochloride on leukocytes count of rats. Different letters refer to significant differences among groups at (P \leq 0.05).

دور الكويرستين الوقائي ضد التأثير السمي للأنيسدين في معايير الدم للجرذان المختبرية (Rattus Norvegicus)

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ألخلاصة

شملت هذه الدراسة ٢٤ ذكرا من الجرذان المختبرية وتم توزيعها على ثلاث مجموعات بواقع ٨ جرذ ذكر لكل مجموعة. مثلت المجموعة الاولى مجموعة السيطرة وفيها تم تغذية الحيوانات على عليقة نموذجية لمدة ١٥ يوما ثم تم قتلها لأجراء الاختبارات اللازمة عليها. المجموعة الثانية تم تغذيتها على عليقة تحتوي على ١٠٠ ملغم\كغم هيدروكلوريد ألأورثو أنيسيدين لمدة ١٥ يوما ثم تم قتلها لأجراء الاختبارات اللازمة. أما المجموعة الثالثة فقد تمت تغذيتها على عليقة تحتوي على ١٠٠ ملغم\كغم هيدروكلوريد ألأنيسيدين + ٨٠ ملغم\كغم كويرستين دايهايدريت لمدة ١٥ يوما ثم تم قتلها لأجراء اللازمة.

اظهرت النتائج أن استخدام هيدروكلوريد الاورثو انيسدين لمدة ١٥ يوما (مجموعة المعاملة الاولى) قد تسبب بانخفاض معنوي في عدد كريات الم الحمراء، تركيز الهيمو غلوبين، نسبة حجم الخلايا المرصوص، عدد الخلايا البيضاء العدلة والخلايا اللمفاوية وقد تسبب ايضا بارتفاع معنوي في عدد الصفيحات الدموية، العدد الكلي لخلايا الدم البيضاء، وعدد الخلايا وحيدة النواة، الخلايا الحمضة والخلايا القعدة مقارنة مع مجموعة السيطرة عند مستوى المعنوية (20.0≥P). لقد بينت النتائج انه عندما تم استخدام الكوبرستين دايهيدريت كعامل وقائي في مجموعة المعاملة الثانية فأنه قد أظهر تأثيرا محسنا للمعايير المتأثرة بالانيسدين حيث تسبب الكوبرستين بارتفاع معنوي في عدد كريات الم الحمراء، تركيز الهيمو غلوبين، نسبة حجم الخلايا المرصوص، عدد الخلايا البيضاء العدلة والخلايا اللمفاوية وقد تسبب ايضا بانخفاض معنوي في عدد الصفيحات الدموية، العدد الكلي لخلايا الدر البيضاء، وعدد الخلايا وحيدة النواة، الخلايا الحمضة والخلايا القعدة مقارنة مع مجموعة المعاملة الاولى عند البيضاء، وعدد الخلايا وحيدة النواة، الخلايا الحمضة والخلايا القعدة مقارنة مع مجموعة المعاملة الاولى عند مستوى المعنوية (20.05). بالنسبة لمعايير الدم الحسابية (مدى توزيع كرات الدم الحمراء، معدل هيمو غلوبين مستوى المعنوية (20.05). بالنسبة لمعايير الدم الحسابية (مدى توزيع كرات الدم الحمراء، معدل هيمو غلوبين مستوى المعنوية معدل تركيز هيمو غلوبين الخليات الحمراء، ومعدل حجم صفيحات الدم في في الاولى عند الكرية، معدل تركيز هيمو غلوبين الخلية، معدل حجم الكريات الحمراء، ومعدل حجم صفيحات الدم) فأنه لم تكن هذالك فروقات معنوية بين جميع المعاملات ماعدا معدل حجم صفيحات الدم ميراد، في هير وكلوريد الاور ثوانيسدين قد تسبب بانخفاض معنوي في معدل حجم صفيحات الدم حيث انه هيدر وكلوريد معراك ومعلي معنوية بين جميع المعاملات ماعدا معدل حجم صفيحات الدم لمجموعة المعاملة الاولى مقار مع موجموعة المعاملة الثانية ومجموعة السيطرة عند مستوى المعنوية (20.05).

REFERENCES

- European Union Official Journal. (2006). Azocolourants (List of aromatic amines). Appendix 8, Pp: 1-3.
- World Health Organization. (1999). Monographs on the Evaluation of the Carcinogenic Risk of Chemical to Humans. *Lyon. Vol.* 73; Pp. 5–6.
- Stabbert, R., Schaffer, K.-H., Biefel, C., and Rustemeier, K. (2003). Analysis of aromatic amines in cigarette smoke. *Rapid. Commun. Mass Spectrom*. 17, 2125-2132.
- Richter, E., Rosler, S., Scherer, G., Gostomzyk, J G., Grubl, A., Kramer, U., and Behrendt, H. (2001). Haemoglobin adducts from aromatic amines in children in relation to area of residence and exposure to environmental tobacco smoke. *Int. Arch. Occup. Environ. Health* 74, 421-428.
- Eva Stocker. (2002). O-ANISIDINE: RISK REDUCTION. Elaboration of a Risk Reduction Strategy under Existing Substances Regulation (EEC) No 793/93, BE-211. Federal Environment Agency, Vienna, Austria. Pp: 1-24.
- Vidhya A, Indira M. (2009). Protective effect of quercetin in the regression of ethanol-induced hepatotoxicity. *Indian J Pharm Sci*; 71: 527-532.
- Paolillo R, Romano Carratelli C, Rizzo A. (2011). Effect of resveratrol and quercetin in experimental infection by Salmonella enteric serovar Typhimurium. *Int Immunopharmacol;* 11: 149-156.

- Pandey KB, Rizvi SIbrahim. (2009). Current understanding of dietary polyphenols and their role in health and disease. *Curr Nutr Food Sci*; 5: 249-263.
- Neha P. Sangai and Ramtej J. Verma. (2012). Quercettin ameliorates bisphenol A-induced toxicity in mice. Acta Poloniae Pharmaceutica - Drug Research, Vol. 69 No. 3 pp. 557-563.
- Tieppo J, Vercelino R, Dias AS, Silva Vaz MF, Silveira TR, Marroni CA. (2007). Evaluation of the protective effects of quercetin in the hepatopulmonary syndrome. *Food Chem Toxicol*; 45: 1140-1146.
- 11. Reutrakul C., Ningnuek N., Pohmakotr M. (2007). Planta Med. 73, 683.
- 12. Inal ME, Akgun A, Kahraman A. (2002). Radioprotective effects of exogenous glutathione against whole-body gamma-ray irradiation: age- and gender-related changes in malondialdehyde levels, superoxide dismutase and catalase activities in rat liver. *Methods Find Exp Clin Pharmacol;* 24:209-12.
- Janis Ott Joslin. (2009). Blood Collection Techniques. Journal of Exotic Pet Medicine, Vol 18, No 2, Pp: 117–139.
- Naiman K, Dračinska H, Martinkova M, Šulc M, Dračinsky M, Kejikova. L, HodekP, Hudeček J, Liberda J, Schmeiser HH, Frei E, Stiborova M. (2008). Redox cyclingin the metabolism of the environmental pollutant and suspected humancarcinogen o-anisidine by rat and rabbit hepatic microsomes. *Chem ResToxicol* 21: 810–821.
- Dračinska H, Mikšanova M, Svobodova M Smrček S, Frei E, Schmeiser HH, Stiborova M. (2006). Oxidative detoxication of carcinogenic 2nitroanisole by human, rat and rabbit cytochrome P450. *Neuro Endocrinol. Lett*, 27: 9–13.
- Neumann H-G, Ambs S, Bitsch A. (1994). The role of non-genotoxic mechanisms in arylamine carcinogenesis. *Environ Health Perspect* 102, Suppl 6.
- J. Michałowicz, W. Duda. (2006). Phenols Sources and Toxicity. Polish J. of Environ. Stud. Vol. 8, No. 3.Pp: 347-362.
- Kohen, R., and Nyska, A. (2002). Oxidation of biological systems: oxidative stress phenomena, antioxidants, redox reactions, and methods for their quantification. *Toxicol. Pathol.* 30, 620–650.

- Adam Antosik1, Magdalena Szejk1, Arkadiusz Gajek2, Halina Malgorzata Zbikowska1. (2012). The Effects of Gamma-irradiation and Quercetin on Human Red Blood Cells. *Electronic International Interdisciplinary Conference*.Pp:563-567.
- S Asgary, GH Naderi, N Askari. (2005).Protective effect of flavonoids against red blood cell hemolysis by free radicals. *Exp Clin Cardiol*; 10(2):88-90.
- Del Principe D, Menichelli A, De Matteis W, Di Giulio S, Giordani M, Savini I, Finazzi-Agro' A. (1991). Hydrogen peroxide is an intermediate in the platelet activation cascade triggered by collagen, but not by thrombin. *Thromb Res*; 62:365–375.
- Pratico' D, Iuliano L, Alessandri C, Camastra C, Violi F. (1993). Polymorphonuclear leukocyte-derived O2-reactive species activate primed platelets in human whole blood. *Am J Physiol*; 264:H1582– H1587.
- Giuseppe Ambrosio , Isabella Tritto , Paolo Golino (1993). Reactive oxygen metabolites and arterial thrombosis. *Cardiovascular Research* 34 1997. 445–452.
- 24. Yao SK, Ober JC, Gonenne A, Clubb FJ, Krishnaswami A, Ferguson JJ, Anderson VH, Gorecki M, Buja LM, Willerson JT. (1993). Activeoxygen species play a role in mediating platelet aggregation and cyclic flow variations in severely stenosed and endothelium-injured coronary arteries. *Circ Res*; 73:952–967.
- 25. A. W. Boots, G. R. M. M. Haenen, and A. Bast. (2008). "Health effects of quercetin: From antioxidant to nutraceutical," *European Journal of Pharmacology*, vol. 585, no. 2-3, pp. 325-337.
- Fiorani M, Accorsi A, Cantoni O. (2003). Human red blood cells as a natural flavonoid reservoir. *Free Radic Res*; 37:1331-8.
- Kitagawa S, Fujisawa H, Sakurai H. (1992). Scavenging effects of dihydric and polyhydric phenols on superoxide anion radicals, studied by electron spin resonance spectrometry. *Chem Pharm Bull*; 40:304-7.
- Lale A, Herbert JM, Augereau JM, Billon M, Leconte M, Gleye J.(1996). Ability of different flavonoids to inhibit the procoagulant activity of adherent human monocytes. *J Nat Prod*; 59:273-6.

- 29. Adriana Carolina Rodrigues Almeida Silva, Breno Fernando Martins de Almeida, Carolina Soares Soeiro, Wagner Luis Ferreira, Valeria Marcal Felix de Lima, Paulo Cesar Ciarlini. (2013). Oxidative stress, superoxide production, and apoptosis of neutrophils in dogs with chronic kidney disease. *The Canadian Journal of Veterinary Research*; 77:136–141.
- Abousalem, 2A. Elgerwi and 3A. El-Mashad. (2012). Biotoxic and Haemotoxic Effects of Air Pollutants at a Benzene Station on Albino Rats. *International Journal of Animal and Veterinary Advances* 4(3): 187-190.
- Huimin J, Qinjun X, Peijun Z, et al. (2000). Impaired GP-91PHOX gene expression and dysfunction of peripheral blood neutrophils in patients maintaining hemodialysis. *Chin Med J*; 113:120–123.